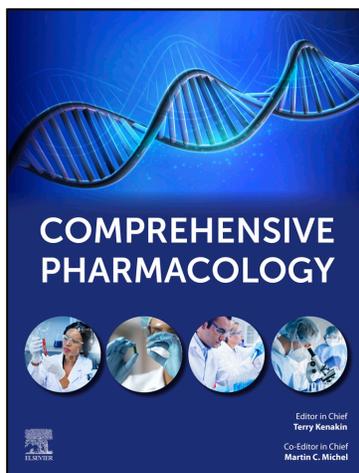


Provided for non-commercial research and educational use.
Not for reproduction, distribution or commercial use.

This article was originally published in *Comprehensive Pharmacology* (PHRM), published by Elsevier, and the attached copy is provided by Elsevier for the author's benefit and for the benefit of the author's institution, for non-commercial research and educational use including without limitation use in instruction at your institution, sending it to specific colleagues who you know, and providing a copy to your institution's administrator.



All other uses, reproduction and distribution, including without limitation commercial reprints, selling or licensing copies or access, or posting on open internet sites, your personal or institution's website or repository, are prohibited. For exceptions, permission may be sought for such use through Elsevier's permissions site at:

<https://www.elsevier.com/about/our-business/policies/copyright/permissions>

From Church, M.K., Frischbutter, S., Kolkhir, P., Maurer, M., 2022. The Pharmacology of Antihistamines. In: Kenakin, T. (Ed.), *Comprehensive Pharmacology*. vol. 5, pp. 515–535. Elsevier. <https://dx.doi.org/10.1016/B978-0-12-820472-6.00072-4>.

ISBN: 9780128204726

Copyright © 2022 Elsevier Inc. All rights reserved

Elsevier

5.28 The Pharmacology of Antihistamines

Martin K. Church^a, Stefan Frischbutter^a, Pavel Kolkhir^{a,b}, and Marcus Maurer^a, ^a Dermatological Allergology, Allergie-Centrum-Charité, Charité - Universitätsmedizin, Berlin, Germany; and ^b Division of Immune-mediated skin diseases, I.M. Sechenov First Moscow State Medical University (Sechenov University), Moscow, Russian Federation

© 2022 Elsevier Inc. All rights reserved.

5.28.1	The effects of histamine in allergic disease	516
5.28.1.1	Histamine H ₁ -receptor effects	516
5.28.1.2	Histamine H ₂ -receptor effects	516
5.28.1.3	Histamine H ₃ -receptor effects	516
5.28.1.4	Histamine H ₄ -receptor effects	517
5.28.2	The discovery and development of antihistamines	517
5.28.2.1	H ₁ -Antihistamines	517
5.28.2.2	H ₂ -, H ₃ - and H ₄ -Antihistamines	518
5.28.3	The histamine H₁-receptor and H₁-antihistamines	518
5.28.3.1	The histamine H ₁ -receptor	518
5.28.3.2	H ₁ -Antihistamines	519
5.28.4	Preclinical pharmacology	519
5.28.4.1	Efficacy	519
5.28.4.2	Speed of onset of action	520
5.28.4.3	Duration of action	521
5.28.4.4	Elimination	522
5.28.4.5	H ₁ -Antihistamines and the central nervous system	522
5.28.4.6	H ₁ -Antihistamines and cardiotoxicity	524
5.28.5	Clinical effectiveness of H₁-antihistamines	525
5.28.5.1	Speed of onset of action and duration of action	525
5.28.5.2	Efficacy	525
5.28.5.3	Anti-inflammatory effects of H ₁ -antihistamines	525
5.28.5.4	H ₁ -Antihistamines in urticaria	525
5.28.5.5	H ₁ -Antihistamines in allergic rhinitis	526
5.28.5.6	H ₂ -Antihistamines in clinical practice	527
5.28.5.7	H ₃ -Antihistamines in clinical practice	527
5.28.6	H₄-Antihistamines in allergic diseases	527
5.28.6.1	JNJ7777120	527
5.28.6.2	JNJ39758979	528
5.28.6.3	Other H ₄ -antihistamines	528
5.28.6.4	H ₄ -Antihistamines in other diseases	529
5.28.7	Conclusion	529
Disclosure of Potential Conflict of Interest		529
References		530

Glossary

Allergic rhinitis Allergic rhinitis is an allergic nasal condition characterized by rhinorrhea and nasal blockage. Rhinorrhea is particularly antihistamine sensitive whereas nasal blockage is less sensitive.

Antihistamines Synthetic compounds that prevent activation of histamine receptors. The older first-generation antihistamines have many unwanted effects including sedation. The newer second-generation antihistamines are more effective and have less unwanted effects.

Histamine An organic amine that has a central role in mediating many of the symptoms of allergic disease, such as each in urticaria and rhinorrhea in allergic rhinitis.

Histamine receptors Cell-surface receptors that are activated by histamine. There are four types, H₁, H₂, H₃ and H₄. In allergic disease H₁ and H₄ are mainly responsible for causing the symptoms. H₃ receptors are primarily in the brain and H₂ mediate gastric acid secretion.

Urticaria Urticaria is an autoallergic skin condition characterized by the occurrence of wheals, angioedema or both. Urticaria can be either spontaneous or inducible. Antihistamines are the primary drug of choice for the treatment of urticaria.

5.28.1 The effects of histamine in allergic disease

In allergic and associated diseases, histamine has diverse effects, both pro-inflammatory and anti-inflammatory, which are determined by both the histamine receptor subtype and the cells stimulated (Thurmond, 2015). Histamine receptors (H_1 -, H_2 -, H_3 - and H_4 -receptors) are characterized by their structure, function, distribution and their affinity to histamine (Leurs et al., 2009; Singh and Jadhav, 2013). The H_1 -receptor drives cellular migration, nociception, vasodilatation and bronchoconstriction (Bakker et al., 2001), whereas the H_2 -receptor modifies gastric acid secretion, airway mucus production and vascular permeability (Seifert et al., 2013). The H_3 -receptor plays an important role in neuro-inflammatory diseases (Singh and Jadhav, 2013). The H_4 -receptor has also been shown to be involved in allergy and inflammation (Thurmond, 2015; Tiligada, 2012). Histamine H_4 -receptor-mediated mast cell activation can regulate a powerful inflammatory cascade by releasing several inflammatory mediators including histamine; these mediators may stimulate the migration of different inflammatory cells to the site of inflammation (Jemima et al., 2014). Likewise, the activation of H_1 -receptor also regulates allergic responses by enhancing the migration of Th2-lymphocytes towards the allergen during lung inflammation (Bryce et al., 2006).

5.28.1.1 Histamine H_1 -receptor effects

The histamine H_1 -receptor is expressed in many tissues and cells, including nerves, respiratory epithelium, endothelial cells, hepatic cells, vascular smooth muscle cells, dendritic cells and lymphocytes (Thurmond et al., 2004; Akdis and Simons, 2006). Histamine activates the H_1 -receptor through $G\alpha_q/11$, which then activates phospholipase C and increases intracellular Ca^{++} levels. As a consequence, histamine elicits the contraction of smooth muscle of the respiratory tract, increases vascular permeability, and induces the production of prostacyclin and platelet activating factor by activating H_1 -receptors (Simons, 2004). Thus, almost all immediate hypersensitivity reactions, including symptoms observed in the skin, such as erythema, pruritus, and edema, may be elicited by the activation of the H_1 -receptor (Schaefer et al., 1999).

Activation of the H_1 -receptor can also enhance both Th1- and Th2-type immune responses (Jutel et al., 2001). Similarly, Bryce et al. (2006) demonstrated that allergen-challenged H_1 -receptor-deficient mice had attenuated lung allergic responses.

In addition, IL-3 activation can increase H_1 -receptor expression on Th1 cells (Horio et al., 2010; Oсна et al., 2001), and histamine can enhance B cell proliferation, which is absent in H_1 -receptor-deficient mice (Banu and Watanabe, 1999). The role of H_1 -receptor activation in asthma is further corroborated by observations showing that use of H_1 -antihistamines can significantly decrease asthma symptoms and improve pulmonary function in persistent asthma (Baena-Cagnani et al., 2003; Simons, 1999b, 2004).

The H_1 -receptor is also expressed in dermal dendritic cells and keratinocytes in the skin tissue, and histamine increases the proliferation of murine epidermal keratinocytes *in situ* (Maurer et al., 1997) and it increases the nerve growth factor production via the H_1 -receptor in human keratinocytes (Kanda and Watanabe, 2003). The secretion of nerve growth factor is caused by the phosphorylation of protein kinase C, extracellular signal-regulated kinases and the activation of activator protein-1 resulting from H_1 -receptor stimulation. Similarly, histamine, acting via the H_1 -receptor, has also been shown to enhance the production of chemokines, such as GM-CSF, RANTES, and MCP-1 in IFN- γ -stimulated keratinocytes. It also up regulates the antigen-presenting capability of dendritic cells, and leads to Th1 polarization through the H_1 -receptor (Giustizieri et al., 2004).

Histamine induces IL-31 production, which plays an important and crucial role in pruritus and skin barrier function in allergic dermatitis (Gutzmer et al., 2009). Administration of an H_1 -antihistamine decreased IL-31 levels in the serum of atopic dermatitis patients (Mommert et al., 2012). These data, taken together, suggest that H_1 -receptor activation by histamine has the ability to induce various symptoms related with allergic and other inflammatory diseases including skin diseases such as chronic urticaria, atopic dermatitis, chronic prurigo, and chronic pruritus.

5.28.1.2 Histamine H_2 -receptor effects

The $G_{\alpha s}$ -coupled H_2 -receptor is highly expressed in various cells and tissues, such as B cells, T cells, dendritic cells, gastric parietal cells, smooth muscle cells, and the brain and cardiac tissues. Activation of the receptor can induce airway mucus production, vascular permeability and secretion of gastric acid (Smit et al., 1996). The H_2 -receptor is importantly involved in relaxation of the airways, uterus and smooth muscle cells in the blood vessels. Moreover, the H_2 -receptor is involved in the activation of the immune system, such as Th1 cytokine production, reduction of basophil degranulation, T-cell proliferation and antibody synthesis (Meiler et al., 2008; Lichtenstein and Gillespie, 1975).

5.28.1.3 Histamine H_3 -receptor effects

The H_3 -receptor is coupled to $G_{\alpha i/o}$ and exclusively expressed in neurones. It is important for homeostatic regulation of energy levels, sleep-wake cycle, cognition and inflammation (Dimitriadou et al., 1994). Studies suggest that the H_3 -receptor can lead to an increase in severity of neuro-inflammatory diseases and can enhance the expression of IFN-inducible protein 10, MIP 2 and

CXCR3 in T cells (Teuscher et al., 2007). The H₃-receptor has also been associated with rhinitis (Lieberman, 2011). This is likely because it is expressed on presynaptic nerves in the peripheral sympathetic adrenergic system and also on nasal sub-mucosal glands. Stimulation of H₃-receptor suppressed norepinephrine release at pre-synaptic nerve endings and stimulated nasal sub-mucosal gland secretion (Suzuki et al., 2008).

5.28.1.4 Histamine H₄-receptor effects

Many cells express H₄-receptors including neurons, fibroblasts, spleen, intestinal epithelia, lung, synovial tissue and immune cells such as eosinophils, neutrophils, basophils and mast cells (Nicoud et al., 2019; Thangam et al., 2018; Lippert et al., 2004). Signal pathways include a decrease in cAMP accumulation through inhibition of adenylyl cyclase, increase in Ca²⁺ mobilization via IP₃, activation of MAPK/ERK, PI3K and SAPK/JNK and inhibition of TGF-β1 signaling pathways and recruitment of β-arrestin (Nicoud et al., 2019).

The H₄-receptor contributes to pain control (Coruzzi et al., 2007; Sanna et al., 2020), has a potential role in cancer progression (Nicoud et al., 2019; Schirmer et al., 2020). It also modulates innate and adaptive immune responses (Nicoud et al., 2019) mediating cell migration and cytokine production (Peng et al., 2019). For example, activation of H₄-receptors has been shown to induce chemotaxis of mast cells, eosinophils and dendritic cells, induction of migration of regulatory T cells and release of mediators including IL-4, -5, -13, -6, -1b, -10, -8 and MCP-1 (Wang et al., 2019; Nicoud et al., 2019).

Histamine H₄-receptor is thought to be involved in allergic diseases and pruritus development and, consequently, is considered as a novel drug target for the treatment of allergy. H₄-receptors are expressed on mast cells, eosinophils, basophils, dendritic and natural killer cells (Thangam et al., 2018; Lippert et al., 2004; Jemima et al., 2014).

Mast cell chemotaxis into sites of allergic inflammation has been shown to be mediated, at least in part, by histamine-activated H₄-receptors. This is prevented by the dual H₃/H₄-antihistamine thioperamide, but not by H₁- or H₂-antihistamines (Hofstra et al., 2003). In another study, JNJ777120, a selective histamine H₄-antihistamine, effectively reversed the chemotaxis of human lung mast cells induced by the H₄-receptor agonist, JNJ28610244 (Kay et al., 2018).

In human mast cells, H₄-receptor activation leads to intracellular Ca²⁺ release, degranulation and the production of Th2 and pro-inflammatory cytokines, e.g. IL-13 and RANTES (Ebenezer et al., 2017, 2018). H₄-receptor gene silencing and pretreatment with JNJ777120 decreased phosphorylation of SAPK/JNK and inhibited the intracellular Ca²⁺ release and degranulation induced by H₄-antihistamines in the human mast cell line-1 (Ebenezer et al., 2018).

In addition, the H₄-receptor may play a role in IgE-induced FcεRI upregulation and in the sensitization phase of mast cell degranulation. In particular, UR-63325, a selective H₄-receptor antagonist/inverse agonist, inhibited antigen-dependent degranulation of murine bone marrow-derived mast cells and IgE-induced FcεRI upregulation (Mirzahosseini et al., 2013).

Furthermore, H₄-receptor is crucial for the histamine-induced adhesion of eosinophils to endothelial cells. For example, Grosicki and colleagues observed concentration-dependent increase and decrease in the number of eosinophils that adhere to endothelium after activation of H₄-receptors with H₄-agonists and blocking with H₄-antihistamines, respectively (Grosicki et al., 2016).

The H₄-receptor has been reported to be highly expressed on keratinocytes from patients with atopic dermatitis. The stimulation of the H₄-receptor increased proliferation of keratinocytes, which was diminished with JNJ777120 (Glatzer et al., 2013). In a mouse model of atopic dermatitis, H₄-receptor-deficient mice showed decreases in skin lesions, influx of inflammatory cells and epidermal hyperproliferation at lesional skin sites, and reduced levels of ovalbumin-specific IgE (Rossbach et al., 2016).

Histamine and the selective H₄-agonist, 4-methylhistamine, caused scratching responses in mice, which were almost completely attenuated in H₄-receptor knockout mice or by oral administration of JNJ 777120 (Dunford et al., 2007). Itch induction may be associated with the intracellular signaling mechanism via the ion channel TRPV1 upon H₄-receptor activation on the dorsal root ganglion neurons (Jian et al., 2015).

Finally, Ohsawa et al. reported that bone marrow-derived basophils preferentially express H₄-receptor mRNA and that the histamine-induced chemotaxis of basophils was blocked by an H₄-antihistamine, but not by an H₁-antihistamine (Ohsawa and Hirasawa, 2012). In a mouse model of allergic rhinitis, mice with H₄R-deficient basophils, H₄-antihistamine-treated wild-type mice, and wild-type mice depleted of basophils failed to develop early or late phase nasal responses following allergen sensitization and challenge (Shiraishi et al., 2013).

5.28.2 The discovery and development of antihistamines

5.28.2.1 H₁-Antihistamines

To understand the strengths and weaknesses of antihistamines, and particularly, H₁-antihistamines, it is necessary to appreciate how they were developed in the 1930s. In his review about his own work (Bovet, 1950), Daniel Bovet wrote "Three naturally occurring amines, acetylcholine, epinephrine, and histamine, may be grouped together because they have a similar chemical structure, are all present in the body fluids, and exert characteristically strong pharmacologic activities. There are alkaloids that interfere with the effects of acetylcholine. Similarly, there are sympatholytic poisons that neutralize or reverse the effects of epinephrine. It seemed possible to me, therefore, that some substance might exist which exerts a specific antagonism toward histamine." It was against this background that Bovet, who was looking for antagonists of acetylcholine, asked his student, Anne-Marie Staub, to test some of these compounds against histamine. Anne-Marie Staub, who was preparing her doctorate thesis in his laboratory, used three

types of laboratory methods for the evaluation of the degree of activity of the various compounds (Bovet, 1950). In the first test, they determined the action against the lethal effects of histamine in guinea pigs. This test they believed to be 'perfectly specific.' In the second test, they determined the protection against histamine administered in the form of an aerosol. Here, they believed that symptoms similar to asthma were produced. In the third test for determining antihistaminic activity, which they believed to be the least specific one, they ascertained the effect of compounds on histamine-induced spasm of the isolated guinea pig ileum. These tests led to the discovery of the first H₁-antihistamine, thymoxyethyldiethylamine (929 F) in 1937 (Staub, 1937).

Although thymoxyethyldiethylamine was too toxic for use in humans, it opened the door for the introduction of the 1st generation H₁-antihistamines into the clinic. These included antergan in 1942 (Halpern, 1942), followed by diphenhydramine in 1945 (Loew et al., 1946) and chlorpheniramine, brompheniramine and promethazine later the same decade (Emanuel, 1999). It should be remembered, however, that these 1st generation H₁-antihistamines (fgAHs) derive from the same chemical stem as cholinergic muscarinic antagonists. Also, early tranquilizers, anti-psychotics, antihypertensive and local anesthetic agents were also developed from this stem. It is hardly surprising, therefore, that fgAHs have poor receptor selectivity and often interact with receptors of other biologically active amines causing anti-muscarinic, anti- α -adrenergic and anti-serotonin effects (Church et al., 2010; Yanai, 2012).

5.28.2.2 H₂-, H₃- and H₄-Antihistamines

While the classical antihistamines discovered in the 1940s and 1950s, such as mepyramine, suppressed the effects of histamine in contracting the smooth muscle of various organs, such as the gut and bronchi, actions such as increased gastric acid secretion, increased heart rate and contraction of the rat uterus were 'mepyramine resistant.' This observation led Ash and Schild (1966) to hypothesize that histamine exerted its effects through more than one receptor and led them to define the classical 'mepyramine sensitive' receptor as the histamine H₁-receptor.

In order to find drugs that antagonized the 'mepyramine resistant' effects of histamine, particularly its stimulation of gastric acid secretion, Sir James Black and his colleagues at The Research Institute of Smith Kline and French Laboratories began in 1964 to synthesize and test over 700 compounds which were closely related to the structure of histamine (Black et al., 1972). This work led to the definition of the histamine H₂-receptor and the discovery of burimamide and cimetidine (Brimblecombe et al., 1975), the forerunners of the H₂-antihistamines for the treatment of gastric ulcers.

Work by Jean-Charles Schwartz and colleagues in Paris in the early 1980s demonstrated that histamine was not only a transmitter in the periphery but also in the brain (Schwartz et al., 1980). However, they also realized that histamine, like other neurotransmitters, may modulate its own release through a presynaptic receptor, an action they suggested to be mediated by a class of receptor, designated as H₃, which was pharmacologically distinct from the previously characterized histamine H₁- and H₂-receptors (Arrang et al., 1983). This discovery and the realization that histamine H₃-receptors are located almost exclusively in the brain has led to the investigation of H₃-antihistamines for the treatment of cognitive disorders and Alzheimer's disease (Brioni et al., 2011).

Using the DNA sequence of the histamine H₃-receptor, several research groups independently identified a previously unexplored G-protein coupled receptor sequence in the human genome as a new histamine receptor, the histamine H₄-receptor (Leurs et al., 2009).

The histamine H₄-receptor was identified in data from human genome databases independently by different scientific groups at the beginning of the millennium. It is a G α i/o-coupled receptor encoded by the H₄-receptor gene (Nicoud et al., 2019; Oda et al., 2000). The H₄-receptor consists of 390 amino acids, localized on 18q11.2 chromosome and has about 40% homology with the human histamine H₃-receptor (Nakamura et al., 2000; Oda et al., 2000; Nicoud et al., 2019). The identification of histamine H₄-receptors on hemopoietic cells, eosinophils, mast cells and dendritic cells and its potential role in chemotaxis and activation of these cells has stimulated widespread research into the possible use of H₄-antihistamines in allergic disease (Smits et al., 2009).

5.28.3 The histamine H₁-receptor and H₁-antihistamines

5.28.3.1 The histamine H₁-receptor

The histamine H₁-receptor is a member of the superfamily of G-protein coupled receptors (GPCRs). Physically they are composed of seven transmembrane domains coupling the exterior domains to the intracellular activating mechanism (Fig. 1). In the way that they work, GPCRs may be viewed as 'cellular switches' that exist as an equilibrium between the inactive or 'off' state and the active or 'on' state (Leurs et al., 2002). To stimulate the receptor, histamine cross links domains III and V to stabilize the receptor in its active conformation or 'on' position (Wieland et al., 1999) (Fig. 1). This is a transient event with histamine being rapidly removed. H₁-antihistamines are not structurally related to histamine and do not prevent the binding of histamine but bind to different sites on the receptor and stabilize it in its inactive form. For example, cetirizine cross links sites on transmembrane domains IV and VI to stabilize the receptor in the inactive state and swing the equilibrium to the 'off' position (Gillard et al., 2002) (Fig. 1). Binding times for H₁-antihistamines vary from 25 s for diphenhydramine to 60 and 73 min for fexofenadine and bilastine, respectively (Bosma et al., 2018). Thus, H₁-antihistamines are not receptor antagonists but are inverse agonists in that they produce the opposite effect on the receptor to histamine (Leurs et al., 2002). Consequently, the preferred term to define these drugs is 'H₁-antihistamines' rather than 'histamine antagonists.'

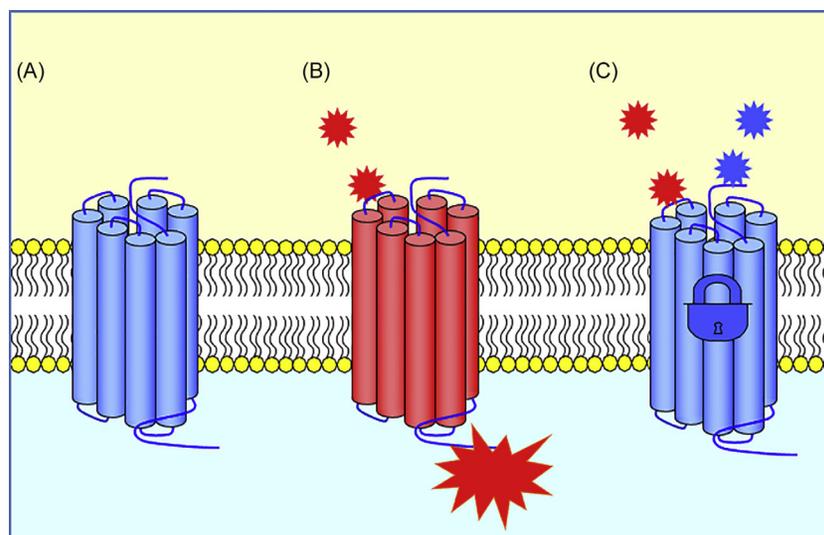


Fig. 1 H₁-antihistamines are inverse agonists not receptor antagonists. (A) unstimulated resting H₁-receptor. (B) Activated H₁-receptor stimulated by histamine (red stars). (C) H₁-receptor locked in the inactive configuration by an H₁-antihistamine (blue stars). Note that histamine may still bind but cannot stimulate. Data from Leurs R, Church MK, Tagliatela M. (2002) H₁-antihistamines: inverse agonism, anti-inflammatory actions and cardiac effects. *Clinical and Experimental Allergy* 32: 489–498.

5.28.3.2 H₁-Antihistamines

There are many efficacious and safe H₁-antihistamines on the market for the treatment of chronic urticaria and allergic rhinitis. It is not the purpose of this chapter to assess and compare individual drugs. However, if readers wish to read the properties of individual drugs, the following reviews are recommended: acrivastine (Bojkowski et al., 1989; Brogden and McTavish, 1991; Gibson et al., 1989), bilastine (Jauregui et al., 2013; Church and Labeaga, 2017; Church, 2011), cetirizine (Spencer et al., 1993; Curran et al., 2004; Broide, 1995), desloratadine (Yuan et al., 2017; Ring et al., 2001; Bachert and Maurer, 2010), ebastine (Van Cauwenberge et al., 2004; Sastre, 2008), fexofenadine (Simpson and Jarvis, 2000; Kawashima et al., 2002), levocetirizine (Singh-Franco et al., 2009; Nettis et al., 2008; Kapp and Wedi, 2004; Ducharme and Weinberg, 2009), loratadine (Monroe, 1996), mizolastine (Lebrun-Vignes et al., 2001; Simons, 1999c), rupatadine (Nettis et al., 2013; Mullol et al., 2008; Metz and Maurer, 2011; Gonzalez-Nunez et al., 2016).

5.28.4 Preclinical pharmacology

5.28.4.1 Efficacy

The efficacy of an H₁-antihistamine is determined by 2 factors: the affinity of the drug for H₁-receptors (absolute potency) and the concentration of the drug at the sites of the H₁-receptors.

The affinity of an H₁-antihistamine for H₁-receptors is determined in preclinical *in vitro* studies. Unfortunately, different methods and target tissues yield differing results so only data from single studies can be compared. In one study using cloned human histamine H₁-receptors, the binding affinities (K_i) in order of potency were; desloratadine 0.4 nM, levocetirizine 2 nM, terfenadine 2 nM, cetirizine 6 nM, fexofenadine 10 nM, and loratadine 16 nM (Gillard et al., 2003). In human H₁-receptors cloned in Chinese hamster ovary cells the affinities were: desloratadine 0.87 nM, chlorpheniramine 2 nM, hydroxyzine 10 nM, terfenadine 40 nM, cetirizine 47 nM, loratadine 138 nM, and fexofenadine 175 nM, (Limon and Kockler, 2003). Finally, in guinea-pig cerebellum, binding affinities were: fexofenadine 6.62 nM, cetirizine 6.85 nM, bilastine 7.37 nM, and ketotifen 9.34 nM (Corcostegui et al., 2005). Although these are often considered to be fixed values, they may be influenced by temperature and pH, and therefore, they can differ in physiological and pathological conditions. For example, in inflammation the pH of the tissues is reduced (Hunt et al., 2000) from 7.4 to 5.8, leading to a 2- to 5-fold increase in the affinity of fexofenadine and levocetirizine for H₁-receptors but no change in the affinity of desloratadine (Gillard and Chatelain, 2006).

In addition to receptor affinity, the concentration of the drug at H₁-receptors *in vivo* is also crucially important. H₁-receptors are situated on the cellular membranes of cells, including vascular and airways smooth muscle, mucous glands, and sensory nerves, all of which are surrounded by extracellular fluid. Many factors affect the concentration of free drug in this compartment, absorption, volume of distribution and plasma protein binding being of prime importance.

Most H₁-antihistamines are absorbed passively into the blood from the intestine, the exceptions being bilastine and fexofenadine as will be described later. Peak plasma levels of passively absorbed drugs occur at around 2–4 h (Geha and Meltzer, 2001; Simons, 2004). Bilastine, and to a lesser extent fexofenadine, have a more rapid uptake as they are substrates for an organic anion

transporting polypeptide, OATP1A2 (Fig. 2) (Shimizu et al., 2005; Lucero et al., 2012; Church and Labeaga, 2017; Cvetkovic et al., 1999; Russell et al., 1998; Tannergren et al., 2003). The role of this transporter is supported by its inhibition by grapefruit juice (Dresser et al., 2005; Crean et al., 2007; Akamine et al., 2015). The mean oral bioavailability of bilastine has been estimated to be around 61% in healthy human volunteers (Lucero et al., 2012), while that of fexofenadine is 30% (Lappin et al., 2010), showing that the affinity for anion pump is stronger with bilastine.

Second is the extent of plasma protein binding which, with H₁-antihistamines, is high, varying from 98% for mizolastine (Simons, 1999c), 96% for cetirizine (Chen, 2008), 90% for levocetirizine (Gillard et al., 2005), 84–90% for bilastine (Church, 2011; Jauregizar et al., 2009), 65% for desloratadine (Gillard et al., 2005) and 60–70% for fexofenadine (Molimard et al., 2004).

Third, and probably most influential, is the apparent volume of distribution, which determines the plasma concentration of a drug after complete body distribution. The apparent volume of distribution is limited for levocetirizine (0.4 L/kg), larger for bilastine (1.29 L/kg) (Church, 2011, Jauregizar et al., 2009) and fexofenadine (5.4–5.8 L/kg) and particularly large for desloratadine (~49 L/kg) (Molimard et al., 2004). The large apparent volume of distribution of desloratadine is primarily due to its extensive intracellular uptake. In the study of Gillard and colleagues (Gillard et al., 2005), the 4-hour plasma concentrations of levocetirizine, desloratadine, and fexofenadine were 28, 1, and 174 nM, respectively.

Because data on the concentrations of H₁-antihistamines in relevant extracellular fluids is generally lacking, the best indirect estimate of efficacy is obtained by calculating receptor occupancy from knowledge of absolute potency and peak drug concentrations in the plasma, usually at ~4 h after a single oral dose (Gillard et al., 2005). This calculation of receptor occupancy after single oral doses of drug shows values of 95%, 90%, and 71% for fexofenadine, levocetirizine, and desloratadine, respectively, indicating that they are all very effective H₁-antihistamines. Receptor occupancy for these drugs appears to correlate with pharmacodynamic activity in skin wheal and flare studies and with efficacy in allergen challenge chamber studies (Gillman et al., 2009; Popov et al., 2006), but is it relevant in clinical practice? Studies in allergic rhinitis suggest that the above 3 drugs are of similar effectiveness (Bachert, 2009; Berger et al., 2006). However, in chronic urticaria, in which local histamine concentrations are high, the differences do seem to be important. For example, in head to head studies in this condition levocetirizine appears significantly more effective than desloratadine (Potter et al., 2009; Staevska et al., 2010).

5.28.4.2 Speed of onset of action

The speed of onset of action of a drug is often equated to the rate of its oral absorption and its duration of action by its plasma concentration. However, this is not strictly correct as the time for a drug to diffuse into the extravascular space to produce a maximal clinical effect is critical. This is seen in Fig. 3, which shows the inhibition of the histamine-induced flare response plotted against the concentration of free drug in the plasma. In this study in children (Simons et al., 2007), plasma concentrations of drug were near maximum by 30 min and yet it took an additional 1.5 h for the drug to diffuse into the extravascular space to produce a maximal

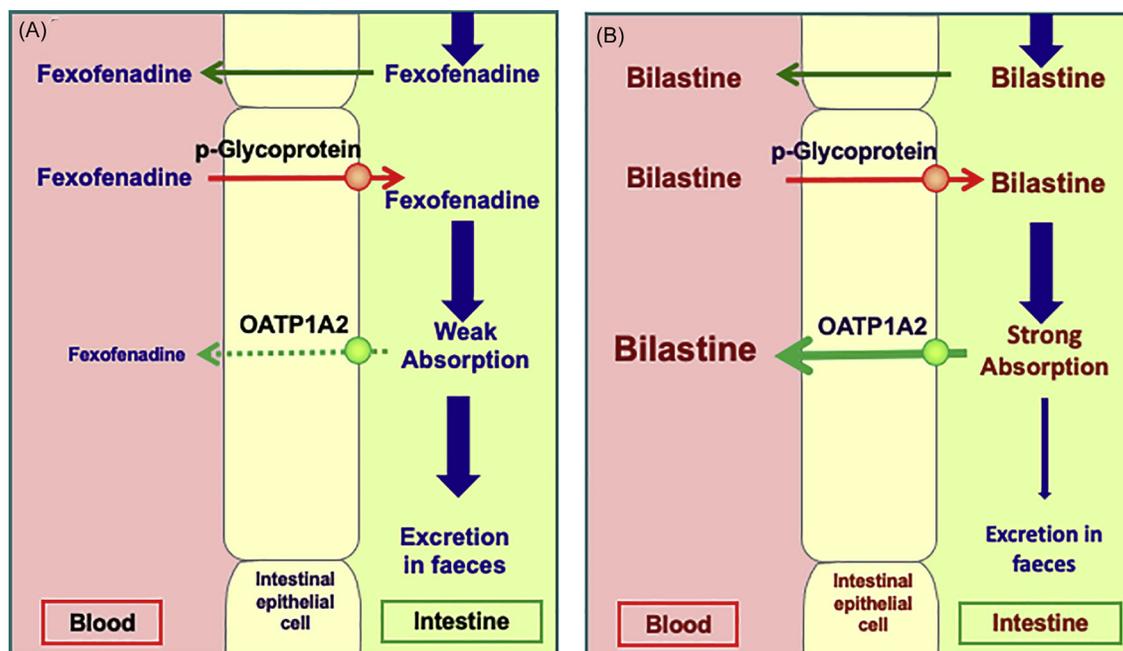


Fig. 2 The roles of p-glycoprotein in causing active excretion of A fexofenadine and B bilastine from the blood into the intestine and of OATP1A2 in causing their reuptake from the intestine into the blood (Cvetkovic et al., 1999). Reproduced from Church MK and Labeaga L (2017) Bilastine: A new H₁-antihistamine with an optimal profile for up dosing in urticaria. *Journal of the European Academy of Dermatology and Venereology* 31: 1447–1452.

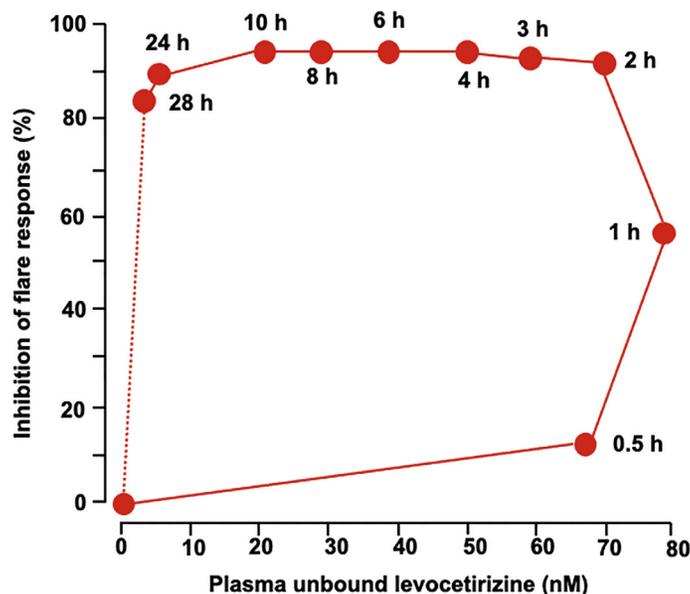


Fig. 3 Hysteresis loop of the inhibition of the histamine-induced flare response plotted against the plasma concentration of unbound levocetirizine after administration of a single 5-mg dose to children. Redrawn from Simons KJ, Benedetti MS, Simons FE, Gillard M, and Baltes E (2007). Relevance of H₁-receptor occupancy to H₁-antihistamine dosing in children. *The Journal of Allergy and Clinical Immunology* 119: 1551–1554.

clinical effect. In adults, the maximal inhibition of the flare response is usually ~4 h for levocetirizine, fexofenadine, and desloratadine (Grant et al., 2002; Purohit et al., 2003, 2004), but may be longer for drugs, such as loratadine and ebastine, which require metabolism to produce their active moiety (Grant et al., 2002). As mentioned earlier, the absorption of bilastine is facilitated by an organic anion transporting polypeptide. As a consequence, bilastine is more rapidly absorbed than passively absorbed drugs (Church, 2011; Church and Labeaga, 2017). An example of the rates of onset of bilastine and levocetirizine is shown in Fig. 4.

5.28.4.3 Duration of action

The duration of action of H₁-antihistamines is not related to their plasma concentration. This is shown clearly in Fig. 3 in which the duration of action of levocetirizine in inhibiting the histamine-induced flare response is much longer than would be predicted from its plasma concentration (Simons et al., 2007). This is presumably due to “trapping” of the drug by its strong and long-lasting binding to histamine H₁-receptors (Gillard et al., 2002). Although less active in the wheal and flare test, desloratadine has a similarly

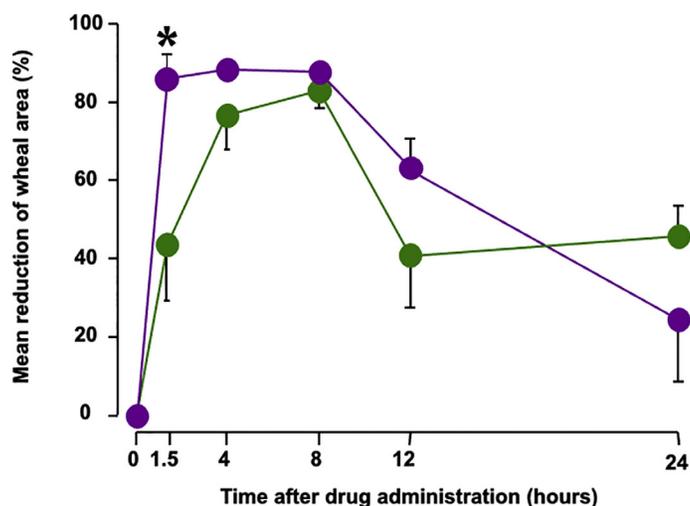


Fig. 4 Inhibition of histamine-induced wheal responses by bilastine 20 mg (purple circles) and cetirizine 10 mg (green circles). *Indicates significance ($P < 0.02$) of difference between bilastine and cetirizine at 1.5 h. Each point is the mean \pm SEM of measurements in 12 healthy volunteers. Reproduced from Church MK and Labeaga L (2017) Bilastine: A new H₁-antihistamine with an optimal profile for up dosing in urticaria. *Journal of the European Academy of Dermatology and Venereology* 31: 1447–1452.

long duration of action (Purohit et al., 2003). However, the duration of action of fexofenadine, calculated in the study of Purohit et al. (2001) as the time for the wheal to be inhibited by at least 70%, is less prolonged, being 8.5 h for 120 mg fexofenadine compared with 19 h for cetirizine. The primary reason for the shorter duration of action of fexofenadine is that it is actively secreted into the intestine and urine by p-glycoprotein (Miura and Uno, 2010). Interestingly, the duration of action of bilastine, which is also a substrate for p-glycoprotein, is much longer because of its active uptake by OATP1A2 (Church and Labeaga, 2017) and because bilastine has an especially long residency time on the H₁-receptor (Bosma et al., 2018). Even so, the guidelines recommend that sgAHs should be taken daily and regularly for the treatment of patients with chronic urticaria in order to obtain maximum efficacy (Zuberbier et al., 2018).

5.28.4.4 Elimination

The metabolism and elimination of H₁-antihistamines have been extensively reviewed elsewhere (Molimard et al., 2004; Devillier et al., 2008; Church and Labeaga, 2017). Cetirizine and levocetirizine are not metabolized and are excreted primarily unchanged in the urine (Molimard et al., 2004). Desloratadine undergoes extensive metabolism in the liver. Although this gives the potential for drug-drug interactions, no significant interactions have been reported (Devilleier et al., 2008). Fexofenadine, which is also minimally metabolized, is excreted primarily in the feces after its active secretion into the intestine under the influence of active drug-transporting molecule, p-glycoprotein (Devilleier et al., 2008). This gives the potential for interactions with agents such as grapefruit juice and St John's Wort, which inhibit these transporters. Although plasma concentrations of fexofenadine may be increased by these agents, no significant resulting adverse reactions have been reported (Devilleier et al., 2008).

5.28.4.5 H₁-Antihistamines and the central nervous system

Perhaps the greatest drawback of fgAHs is their ability to cross blood-brain barrier and interfere with histaminergic transmission. Histamine is an important neuromediator in the human brain, which contains approximately 64,000 histamine-producing neurons, emanating from the tuberomammillary nucleus (Haas and Panula, 2003). Stimulation of H₁-receptors in the CNS increases arousal in the circadian sleep/wake cycle, reinforces learning and memory, and has roles in fluid balance, suppression of feeding, control of body temperature, control of the cardiovascular system and mediation of stress-triggered release of ACTH and β -endorphin from the pituitary gland (Brown et al., 2001). It is not surprising then that fgAHs, such as chlorpheniramine, diphenhydramine, hydroxyzine and ketotifen, which, even when given at licensed doses, occupy more than 50% of brain H₁-receptors, interfere with all of these processes (Fig. 5) (Yanai et al., 2011).

Physiologically, the release of histamine during the day causes arousal, whereas its decreased production at night results in a passive reduction of the arousal response. When taken during the day, fgAHs, even at the manufacturers' recommended doses, frequently cause daytime somnolence, sedation, drowsiness, fatigue and impaired concentration and memory (Simons, 2004; Juniper et al., 2005). When taken at night, fgAHs increase the latency to the onset of rapid eye movement (REM) sleep and reduce the duration of REM sleep (Fig. 6) (Adam and Oswald, 1986; Boyle et al., 2006; Rojas-Zamorano et al., 2009; Church et al., 2010). The residual effects of poor sleep, including impairment of attention, vigilance, working memory and sensory-motor performance, are still present the next morning (Boyle et al., 2006; Kay et al., 1997). This is especially problematic with drugs with a long half-life, such as chlorpheniramine (21–27 h), hydroxyzine (20–25 h) and promethazine (16–19 h).

In adults, first-generation H₁-antihistamines in doses commonly recommended for the treatment of allergic disorders frequently lead to daytime somnolence, sedation, drowsiness, fatigue and impaired concentration and memory (Simons, 1999a, 1994; Juniper et al., 2005; Church et al., 2010). The incidence of subjectively reported somnolence has been reported to vary from 40% with chlorpheniramine or brompheniramine to 80% after hydroxyzine (Druce et al., 1998). However, lack of drowsiness does not mean that an individual is able to drive a vehicle without impairment because subjective somnolence and impairment of the ability to perform tasks are not necessarily correlated, some individuals denying subjective adverse effects despite objective evidence of impairment, while others complain of somnolence and yet are able to perform psychomotor tests adequately (Warren et al., 1981; Simons et al., 1995).

In children, the detrimental CNS effects of fgAHs on learning and examination performance are again well established (Church et al., 2010). Allergic rhinitis reduces learning ability in children and is associated with poor examination performance in teenagers. This situation is exacerbated by fgAHs (Vuurman et al., 1993, 1996; Scadding, 2008). In an analysis of 1834 teenage students in the UK taking national examinations, those with untreated allergic rhinitis were 40% more likely to drop one or more grades compared with healthy teenagers. However, if they took a fgAH this figure increased to 70% (Walker et al., 2007).

A major advance in antihistamine development occurred in the 1980s with the introduction of sgAHs, including loratadine, desloratadine, cetirizine, levocetirizine, ebastine, rupatadine, azelastine and olopatadine, which have high H₁-receptor selectivity, no anti-cholinergic effects, low brain permeability and longer durations of action (Holgate et al., 2003). Unlike fgAHs, sgAHs are amphiphilic in that hydrophilic groups have been introduced into the molecule so that they are always positively or negatively charged and, therefore, have a greatly reduced passage across the blood brain barrier occupying less than 20% of brain H₁-receptors (Fig. 5) (Yanai et al., 2011; Hiraoka et al., 2015). Although sgAHs have a much-reduced brain penetration, they may only be referred to as 'minimally sedating' rather than 'non-sedating.' For example, in a study of patients' perspective of effectiveness and side effects of H₁-antihistamine up-dosing in chronic spontaneous urticaria, more than 20% of patients reported sedation as a side effect of sgAHs (Church et al., 2011).

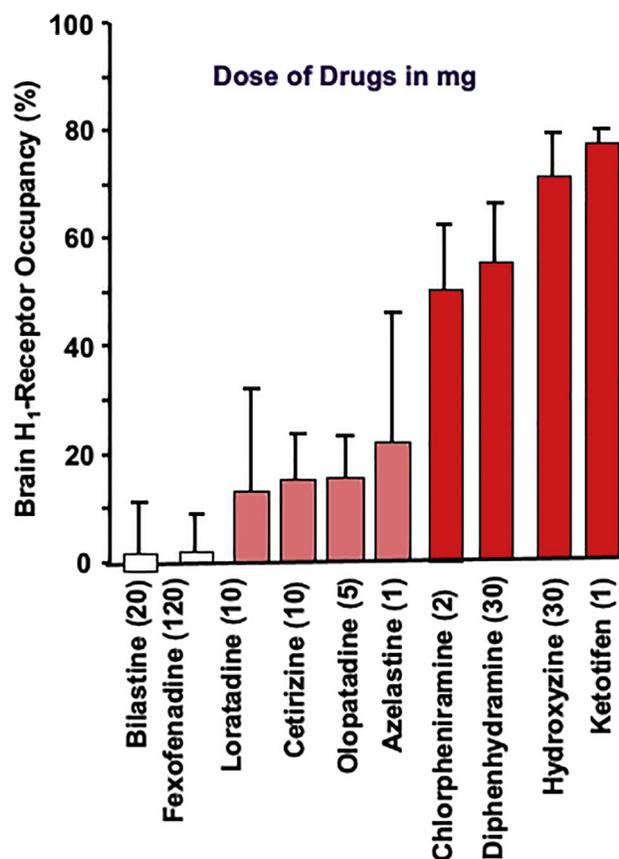


Fig. 5 Percentage occupancy brain H₁-receptors by H₁-antihistamines. First generation agents are colored red and substrates of p-glycoprotein are white. Data from Yanai K, Zhang D, Tashiro M, Yoshikawa T, Naganuma F, Harada R, Nakamura T, Shibuya K, and Okamura N (2011) Positron emission tomography evaluation of sedative properties of antihistamines. *Expert Opinion on Drug Safety* 10: 613–622 and Farre M, Perez-Mana C, Papaseit E, Menoyo E, Perez M, Martin S, Bullich S, Rojas S, Herance JR, Trampal C, Labeaga L, and Valiente R (2014) Bilastine vs. hydroxyzine: occupation of brain histamine H₁-receptors evaluated by positron emission tomography in healthy volunteers. *British Journal of Clinical Pharmacology* 78: 970–980.

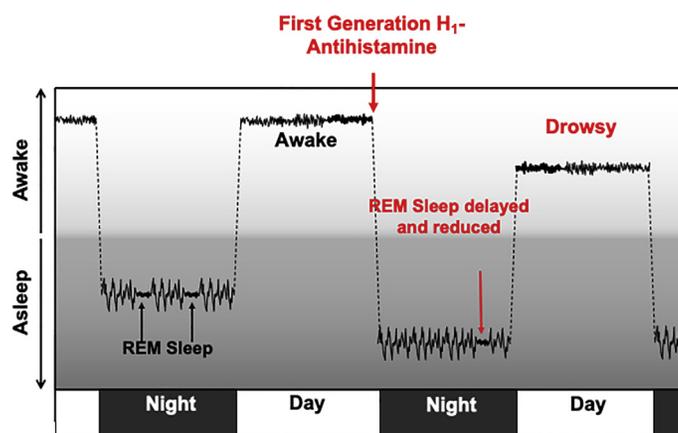


Fig. 6 A theoretical diagram of the sleep/wake cycle and the effects of a first-generation H₁-antihistamine leading to somnolence during the day and abnormal sleep at night. This diagram is reproduced from Church MK, Maurer M, Simons FE, Bindslev-Jensen C, Van Cauwenberge P, Bousquet J, Holgate ST, Zuberbier T, Global A, and Asthma European, N. (2010) Risk of first-generation H₁-antihistamines: A GA(2)LEN position paper. *Allergy* 65: 459–466.

More recently, studies have suggested hydrophilicity alone is not sufficient to keep drugs from entering the brain but that an active efflux transporter in the blood–brain barrier may be involved. The most extensively studied of the active efflux proteins is P-glycoprotein (P-gp), which is known to efflux a wide variety of structurally dissimilar drugs (Chen et al., 2003; Seelig and Landwojtowicz, 2000). *In vitro*, studies of P-gp-mediated efflux from caco-2 cells has shown cetirizine, desloratadine and hydroxyzine to

have weak but significant efflux ratios while that of fexofenadine was much greater (Crowe and Wright, 2012). More recently, similar studies have shown that bilastine also has a high efflux ratio (Burton et al., 2007; Church, 2011). The failure of bilastine and fexofenadine to enter the brain and occupy histamine H_1 -receptors has been confirmed using positron emission tomography (Farre et al., 2014). Thus, these two drugs appear to be truly 'non-sedating' H_1 -antihistamines, and the most likely reason for their lack of brain penetration is that they are actively pumped out of the blood-brain barrier by P-gp (Figs. 5 and 7) (Maurer et al., 2011; Chen et al., 2003; Schinkel, 1999; Montoro et al., 2011; Church, 2011).

5.28.4.6 H_1 -Antihistamines and cardiotoxicity

The effect of H_1 -antihistamines in causing cardiotoxicity has been reviewed in detail recently (Cataldi et al., 2019).

The introduction of the sgAHs in the late 1970s and 1980s brought new and unexpected problems, with an increasing number of reports showing an association between the consumption of astemizole and terfenadine and cardiotoxicity. Both of these are essentially pro-drugs that are metabolized by the cytochrome P450 enzyme, CYP3A4, to their active antihistaminic form. However, it was soon realized that if this metabolism was blocked by the concomitant use of inhibitors of CYP3A4, such as ketoconazole, itraconazole and macrolide antibiotics, or by grapefruit juice, which causes post-translational down-regulation of CYP3A4, then this could cause the prolongation of the QT interval, leading to the appearance of polymorphic ventricular arrhythmias, syncope and even cardiac arrest in susceptible individuals (Leurs et al., 2002).

The mechanism most frequently involved in cardiotoxicity induced by sgAHs is the blockade of hERG (Kv11.1) voltage-gated K^+ channels; these channels contribute to cardiac repolarization by carrying the I_{Kr} current and, therefore, their blockade causes QT prolongation and ultimately torsade de pointes (Roy et al., 1996; Soldovieri et al., 2008; Taglialatela et al., 1998, 1999, 2000; Suessbrich et al., 1996).

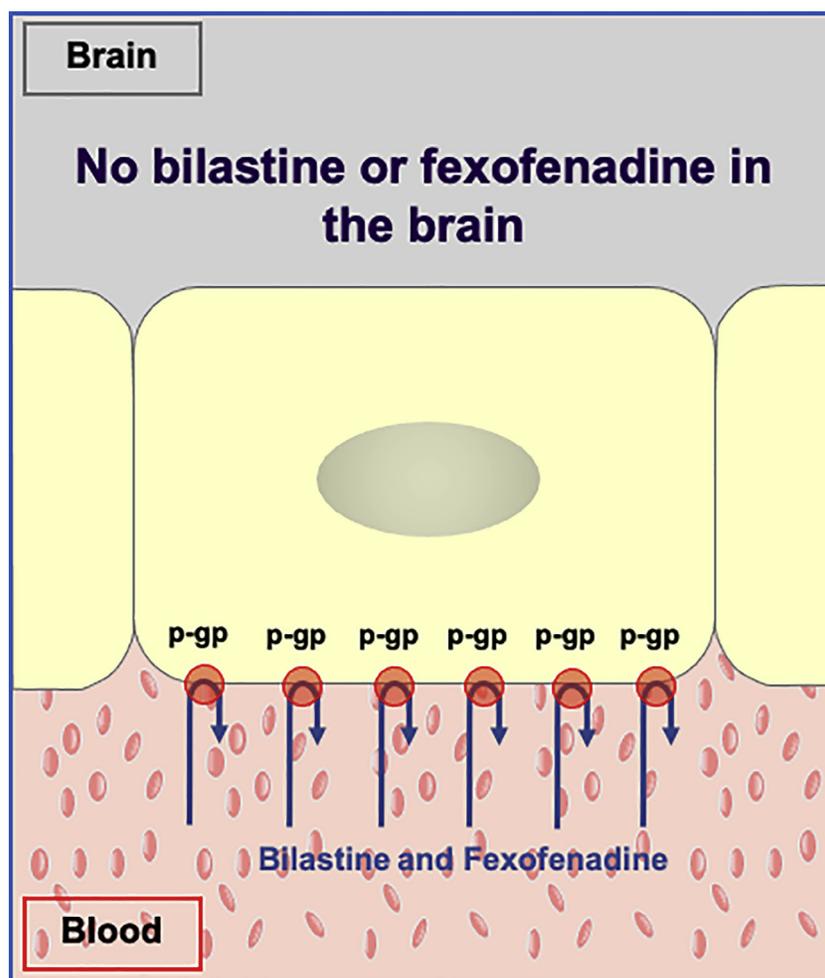


Fig. 7 Diagrammatic representation of the role of p-glycoprotein preventing bilastine and fexofenadine crossing the blood-brain into the brain. Data from Lucero ML, Gonzalo A, Ganza A, Leal N, Soengas I, Iloja E, Gedy S, Jahic M, and Bednarczyk D (2012) Interactions of bilastine, a new oral H_1 antihistamine, with human transporter systems. *Drug and Chemical Toxicology* 35: Supplement 1: 8–17.

Looking at individual H₁-antihistamines, astemizole and terfenadine are no longer approved for use by regulatory agencies in most countries. However, some fgAHs, such as promethazine (Jo et al., 2009), brompheniramine (Park et al., 2008) and chlorpheniramine (Hong and Jo, 2009), may also be associated with a prolonged QTc and cardiac arrhythmias when taken in large doses or overdosed. Today, the concentration of a drug required to produce a half-maximal block of the hERG potassium current (IC₅₀) is used as a surrogate marker for pro-arrhythmic properties of compounds and is the primary test for cardiac safety of drugs (Polak et al., 2009). No clinically significant cardiac effects have been reported for the sgAHs fexofenadine, the metabolite of terfenadine, desloratadine, loratadine, cetirizine, levocetirizine, azelastine, ebastine, mizolastine, rupatadine and bilastine (Ten Eick et al., 2001; DuBuske, 1999; Simons et al., 2003; Hulhoven et al., 2007; Izquierdo et al., 2003; Maurer et al., 2011).

5.28.5 Clinical effectiveness of H₁-antihistamines

5.28.5.1 Speed of onset of action and duration of action

The speed of onset of action of a drug is often equated to the rate of its oral absorption and its duration of action by its plasma concentration at different times. However, this is not strictly correct for drugs which have to diffuse into the extravascular space to produce their maximal clinical effect. In adults, the maximal inhibition of the flare response is usually ~4 h for levocetirizine, fexofenadine and desloratadine (Grant et al., 2002; Denham et al., 2003; Purohit et al., 2004) but may be longer for drugs, such as loratadine and ebastine, which require metabolism to produce their active moiety (Purohit et al., 2004). In contrast, a recent wheal and flare study has suggested that bilastine may have a more rapid onset of action because of its facilitated uptake from the duodenum (Church and Labeaga, 2017).

The duration of action of H₁-antihistamines is also much longer than would be predicted from their plasma concentration and for most is in the vicinity of 24 h (Purohit et al., 2003; Church and Labeaga, 2017). This is presumably due to 'trapping' of the drug by its strong and long lasting binding to histamine H₁-receptors (Gillard et al., 2002). This may be especially so for bilastine, which has an especially long residency time on the H₁-receptor (Bosma et al., 2018). Because it is actively secreted into the bile, intestine and urine by P-glycoprotein (Miura and Uno, 2010), the duration of action of fexofenadine is shorter, around 8¹/₂ h (Purohit et al., 2001), indicating that it may be best given twice daily. In contrast, bilastine, which is also a substrate for p-glycoprotein, has a duration of action of around 24 h. The reason for this difference is that bilastine is also a substrate for OATP1A, an intestinal pump that facilitates its uptake into the bloodstream (Church and Labeaga, 2017; Lucero et al., 2012). Even so, the guidelines recommend that sgAHs should be taken regularly for the treatment of patients with chronic urticaria in order to obtain maximum efficacy (Zuberbier et al., 2018).

5.28.5.2 Efficacy

A question that is asked repeatedly is how the dose of an antihistamine is determined. The answer is that it is a balance between the effectiveness and the unwanted or side effects of a drug. For fgAHs, it is the degree of somnolence they cause that limits the amount of drug that may be given. With most sgAHs, their ability to penetrate the CNS to cause sedation is also a limiting factor. Drugs such as cetirizine, rupatadine and desloratadine may be minimally sedating at licensed doses, but up dosing may cause sedation in susceptible patients. Possible exceptions to this rule are fexofenadine and bilastine which, because they are p-glycoprotein substrates and do not penetrate the CNS, may be up dosed without fear of somnolence (Church and Labeaga, 2017; Maurer et al., 2011; Krause et al., 2013).

5.28.5.3 Anti-inflammatory effects of H₁-antihistamines

The early studies of the anti-inflammatory effects of H₁-antihistamines were reviewed by Leurs et al. (2002); Church, 1999). However, current appreciation of the anti-inflammatory mechanisms mediated through the H₁-receptor stems from the observation by Bakker and colleagues (Bakker et al., 2001) that histamine can activate NF-κB, a transcription factor involved in the synthesis of many pro-inflammatory cytokines and adhesion molecules involved in the initiation and maintenance of allergic inflammation. This is supported by the observation that in its treatment of cold-induced urticaria, bilastine reduced the generation of IL-6 and IL-8 (Krause et al., 2013). The clinical implications of this lie in the ability of H₁-antihistamines to reduce nasal congestion and hyper-reactivity (Bachert, 2009), which result from the sensitization of sensory neurons in the nose by allergic inflammation (Grant et al., 2002). However, as nasal congestion is more slowly relieved than other nasal symptoms (Bachert et al., 2004), continuous rather than on demand therapy with antihistamines is required for its treatment (Canonica et al., 2008).

5.28.5.4 H₁-Antihistamines in urticaria

Most types of urticaria, including chronic spontaneous urticaria (CSU) and the majority of inducible urticarias, are mediated primarily by mast cell-derived histamine (Church et al., 2018), which reaches very high concentrations due to the poor diffusibility of substances in the dermis (Petersen et al., 1997; Church et al., 1997). Urticaria is characterized by short-lived wheals ranging from a few millimeters to several centimeters in diameter, which are accompanied by severe itching that is usually worse in the evening or night-time (Maurer et al., 2011).

The latest EAACI/GA²LEN/EDF/WAO guidelines for the management of urticaria (Zuberbier et al., 2018) recommend that the first-line treatment for urticaria should be a 2nd generation, non-sedating H₁-antihistamine. Furthermore, the guidelines state “We recommend aiming at complete symptom control in urticaria, considering as much as possible the safety and the quality of life of each individual patient.” Because standard licensed doses of H₁-antihistamines are often ineffective in completely relieving symptoms in many patients (Church et al., 2011), the guidelines state “We suggest up dosing sgAHs up to 4-fold in patients with chronic urticaria unresponsive to sgAHs 1-fold” (Fig. 8) (Zuberbier et al., 2018). The guidelines also recommend up dosing with a single antihistamine rather than using different H₁-antihistamines at the same time. If somnolence is a problem, then either fexofenadine or bilastine should be considered. Thus, it is clear that the attributes that clinicians seek when choosing an H₁-antihistamine, are; a rapid onset of action, good efficacy, a long duration of action and freedom from unwanted effects. While some of these attributes may be predicted from pre-clinical and pharmacokinetic studies, it is only in the clinical environment that they may be definitively established (Church and Maurer, 2012).

For children, many clinicians use sedating fgAHs as their first choice assuming that the safety profile of these drugs is better known than that of the newer sgAHs. However, the guidelines make a strong recommendation to discourage the use of fgAHs in infants and children for the reasons stated above. Thus, in children the same first line treatment and up dosing (weight and age adjusted) is recommended as in adults. It should be realized, however, that young children have more body water, as a percentage, than adults. Also, their renal function is fully developed. In contrast, liver enzymes mature more slowly reaching maximum at around 10 years of age. Consequently, in young children, only water-soluble drugs that are excreted renally, such as cetirizine, levocetirizine, fexofenadine and bilastine, should be used.

In elderly patients, fgAHs should also not be used, particularly not in those with dementia as the cumulative use of 1st generation antihistamines with anticholinergic activity is associated with an increased risk for dementia (Gray et al., 2015; Tannenbaum et al., 2012).

5.28.5.5 H₁-Antihistamines in allergic rhinitis

Allergic rhinitis is among the most common diseases globally and usually persists throughout life (Bousquet et al., 2008). The prevalence of self-reported AR has been estimated to be approximately 2% to 25% in children (Asher et al., 2006) and 1% to greater than 40% in adults (Bousquet et al., 2008; Katelaris et al., 2012). The treatment of allergic rhinitis and its impact on asthma (ARIA) guidelines (Brozek et al., 2017) give a comprehensive treatment algorithm.

Clearly, the mast cell histamine-mediated mucus secretion, itching and sneezing are the symptoms that are most sensitive to H₁-antihistamines. H₁-antihistamines are less effective in nasal blockage, which is caused primarily by eosinophil-dominated allergic inflammation, which increases nerve-activated vasodilation in the nose. This symptom is preferentially treated by intranasal corticosteroids (Scadding and Church, 2006).

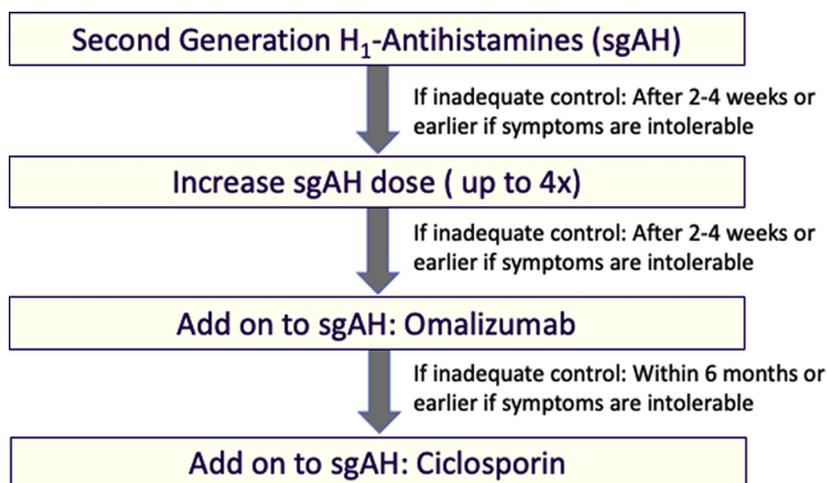


Fig. 8 Recommended treatment algorithm for urticaria from the EAACI/GA²LEN/EDF/WAO guideline for the definition, classification, diagnosis and management of urticaria. The 2017 Revision and Update Zuberbier T, Aberer W, Asero R, Abdul Latiff AH, Baker D, Ballmer-Weber B, Bernstein JA, Bindslev-Jensen C, Brzoza Z, Buense Bedrikow R, Canonica GW, Church MK, Craig T, Danilycheva IV, Dressler C, Ensina LF, Gimenez-Arnau A, Godse K, Goncalo M, Grattan C, Hebert J, Hide M, Kaplan A, Kapp A, Katelaris CH, Kocaturk E, Kulthanan K, Larenas-Linnemann D, Leslie TA, Magerl M, Mathelier-Fusade P, Meshkova RY, Metz M, Nast A, Nettis E, Oude-Elberink H, Rosumeck S, Saini SS, Sanchez-Borges M, Schmid-Grendelmeier P, Staubach P, Sussman G, Toubi E, Vena GA, Vestergaard C, Wedi B, Werner RN, Zhao Z, and Maurer M (2018) The EAACI/GA(2)LEN/EDF/WAO guideline for the definition, classification, diagnosis and management of urticaria. *Allergy* 73: 1393–1414.

5.28.5.6 H₂-Antihistamines in clinical practice

H₂-antihistamines are widely used for the treatment of acid-peptic disease, including duodenal and gastric ulcers and gastroesophageal reflux disease. H₂-antihistamines bind to H₂ receptors of gastric parietal cells and reduce both stimulated and basal gastric acid production and secretion that is induced by histamine. The first of the four H₂-antihistamines currently used was cimetidine, introduced in the late 1970s, followed by ranitidine, famotidine and nizatidine in the 1980s. All are available for oral use and well-absorbed after oral administration. H₂-antihistamines are generally well-tolerated and side effects are uncommon, but in 2019, several ranitidine products were found to contain the carcinogen N-nitrosodimethylamine (NDMA) with subsequent recalls, suspensions and bans in many countries.

The use of H₂-antihistamines, such as cimetidine and ranitidine, in the treatment of urticaria has been controversial with current evidence not supporting their use (Theoharides, 1989). A detailed review of four studies involving a total of 144 participants concluded that although a measure of relief of symptoms of urticaria and rather minimal clinical improvement in some of the participants, the evidence was weak and unreliable (Fedorowicz et al., 2012). The current EAACI/GA²LEN/EDF/WAO urticaria guideline states that H₂-antagonists and dapsone, recommended in the previous versions of the guideline, are now perceived to have little evidence to maintain them as recommendable in the treatment algorithm (Zuberbier et al., 2018).

In allergic rhinitis there appeared to be a small but statistically significant additive effect of H₂-antihistamines to the clinical response to H₁-antihistamines (Carpenter et al., 1983). H₂-antihistamines are not recommended in the allergic rhinitis guidelines (Brozek et al., 2017).

5.28.5.7 H₃-Antihistamines in clinical practice

H₃-antihistamines, as of now, have not found a place in the treatment of allergic diseases. In an early paper, the addition of an H₃-antihistamine to an H₁-antihistamine in the treatment of urticaria was found to be marginally better than the H₁-antihistamine alone (McLeod et al., 2005). However, this was not followed up.

In allergic rhinitis, the combination of fexofenadine, and PF-03654746 caused a reduction in allergen-induced nasal symptoms. The authors suggested that H₃-receptor antagonism might be a novel therapeutic strategy to further explore in patients with allergic rhinitis (Stokes et al., 2012). In another study, prophylactic treatment with the H₃-antagonist JNJ-39220675 relieved allergen-induced nasal congestion by using standard nasal symptom scoring (Barchuk et al., 2013). H₃-antihistamines are not recommended in either the urticaria or allergic rhinitis guidelines (Brozek et al., 2017; Zuberbier et al., 2018).

5.28.6 H₄-Antihistamines in allergic diseases

Several H₄-antihistamines have been developed and some of them are being used in clinical trials for allergic diseases with promising results.

5.28.6.1 JNJ7777120

JNJ7777120 is the first highly selective H₄-antihistamine discovered. It inhibited histamine-induced chemotaxis and calcium influx in mouse bone marrow-derived mast cells, migration of tracheal mast cells from the connective tissue toward the epithelium in mice and neutrophil infiltration in a mouse mast cell-dependent zymosan-induced peritonitis model (Thurmond et al., 2004). JNJ7777120 effects have been extensively evaluated in animal models of allergic skin diseases including atopic dermatitis, contact dermatitis and pruritus.

In a mouse model of atopic dermatitis, treatment with two specific H₄-antihistamines, JNJ7777120 and JNJ28307474, led to a significant reduction in ear edema, inflammation, mast cell and eosinophil infiltration as well as reduction in the levels of several cytokines and chemokines in the ear tissue. Moreover, H₄-antihistamines suppressed lymphocyte proliferation and IL-4, IL-5, and IL-17 levels (Cowden et al., 2010). JNJ7777120 significantly inhibited the production of TH2 chemokines CCL17 and CCL22 in the human monocyte-derived langerhans cells from patients with atopic dermatitis and in antigen-stimulated BMMC (Miyano et al., 2016; Ohsawa and Hirasawa, 2012). NK cell chemotaxis was decreased with JNJ7777120 (Ehling et al., 2016). In another study, combined use of the H₁-antihistamine, cetirizine, and JNJ7777120 decreased serum IgE and levels of IL-4, IL-5 and IL-6 in eczematous lesions (Matsushita et al., 2012).

In addition to its anti-inflammatory effects, JNJ7777120 also significantly inhibited the pruritus in mice, alone (Cowden et al., 2010; Suwa et al., 2011) or in combination with the H₁-antihistamines olopatadine (Ohsawa and Hirasawa, 2012) or cetirizine (Rossbach et al., 2009). In an experimental model of pruritic dermatitis in mice, JNJ7777120 (10 and 30 mg/kg) reduced scratching behavior and ameliorated the skin lesions in a dose-dependent manner and decreased serum IgE levels and numbers of mast cells in skin lesions (Suwa et al., 2011).

JNJ7777120 was effective in chronic allergic contact dermatitis induced by repeated challenge in mice. It suppressed migration of mast cells and eosinophils in skin lesions, decreased serum IgE, IL-4, IL-5 and IL-6 levels and increased levels of interferon-gamma and IL-12 (Seike et al., 2010). JNJ7777120 inhibited extracellular signal-regulated kinase (ERK) activation and spontaneous itching behavior that suggests persistent ERK activation via the H₄R in spinal neurons associated with chronic itch (Huang et al., 2018). In

a mouse model, JNJ7777120 significantly reduced both histamine- and substance P-induced scratching (Yamaura et al., 2009). Pruritus induced by intradermal injection of antigen-specific IgE and antigen stimulation systemically 24 h later was also potently inhibited with JNJ7777120 in H₄R knockout mice (Dunford et al., 2007). Interestingly, the inhibitory effect of JNJ7777120 was greater than those observed with histamine H₁-antihistamines.

Several experimental studies reported the effects of JNJ7777120 in animal models of allergic rhinitis, asthma or peanut allergy.

In an ovalbumin-induced allergic rhinitis rat model, JNJ7777120 significantly improved nasal symptoms, decreased serum levels of IgE, IL-4 and eotaxin in nasal lavage fluid, and increased serum levels of IFN- γ compared with the allergic rhinitis group with no treatment. However, the effect was weak compared with loratadine (Yan et al., 2010). Furthermore, JNJ7777120 dose-dependently inhibited nasal symptoms by single and repeated intranasal and oral administrations in allergic rhinitis in mice. However, the compound decreased serum levels of IgE only by oral administration (Takahashi et al., 2009). JNJ7777120 significantly suppressed nasal symptoms and cough in ovalbumin induced allergic rhinitis which was studied in guinea pigs (Kovacova-Hanusikova et al., 2015).

In ovalbumin-sensitized guinea pigs, JNJ7777120 dose-dependently increased levels of LC-1, reduced allergic asthmatic responses and airway inflammation and lowered levels of PGD₂, LTB₄ and TNF- α in BAL fluid (Somma et al., 2013). In a mouse model of peanut allergy, combined blockade of H₁R (loratadine) and H₄R (JNJ7777120) prevented the development of diarrhea and intestinal inflammation. In addition, the combination of both drugs suppressed dendritic cell functions including antigen-presenting, calcium mobilization and chemotaxis to histamine *in vitro* (Wang et al., 2016). Finally, JNJ7777120 exerted anti-inflammatory and anti-fibrotic effects in an inflammatory mice model of lung fibrosis (Durante et al., 2019).

In summary, JNJ7777120 has been used as the reference tool for studying the functional activity of H₄R in *in vitro* and *in vivo* studies. However, it is considered far from being an ideal antihistamine because of its short *in vivo* half-life and, as reported by Thurmond et al., adrenal toxicity in rats and dogs that prevented its use in clinical studies (Nicoud et al., 2019; Thurmond et al., 2017).

5.28.6.2 JNJ39758979

Another H₄-antihistamine, JNJ39758979, showed a good safety profile in preclinical toxicity studies in rats and monkeys as well as in a phase 1 human volunteer study. After a single oral dose, JNJ39758979 demonstrated a plasma half-life of 124–157 h and dose-dependent inhibition of histamine-induced eosinophil shape change (Thurmond et al., 2014).

In a mouse ovalbumin-induced atopic dermatitis model, mice were treated with the H₁R inverse agonist mepyramine, JNJ39758979 or a combination of both. H₁- and H₄-antihistamines provide synergistic anti-inflammatory effects showing decreased scratching behavior, severity of skin lesions, influx of inflammatory cells, reduced epidermal thickening and low levels of IL-33 in lesional skin. Importantly, drugs given alone did not show this strong anti-inflammatory effect (Köchling et al., 2017).

The safety and efficacy of JNJ39758979 on histamine-induced pruritus in healthy subjects were tested in a randomized clinical trial (RCT) (NCT01068223). Volunteers received a single oral dose of 600 mg JNJ 39758979, 10 mg cetirizine, or placebo. Compared with cetirizine, JNJ 39758979 reduced the AUC of pruritus score but did not demonstrate a significant decrease in wheal or flare. Headache and nausea were reported in some patients (Kollmeier et al., 2014).

A phase 2a RCT of an H₄-antihistamine (JNJ39758979) was carried out in Japanese adults with moderate atopic dermatitis (NCT01497119). Patients were randomized to 300 mg, 100 mg of drug or placebo once daily for 6 weeks. Although the study did not meet the primary end-point, improvements in median EASI were observed with 100 mg and 300 mg versus placebo at week 6. Importantly, two patients (both receiving JNJ39758979 300 mg) showed neutropenia that led to premature study discontinuation (Murata et al., 2015).

The efficacy of JNJ 39758979 has been investigated in adults with persistent asthma but no results have not been reported (NCT00946569).

5.28.6.3 Other H₄-antihistamines

Pre-treatment with compounds A and L markedly reduced the H₄R-mediated intracellular calcium release, phosphorylation of ERK1/2, Akt and NF- κ B, and Th2 cytokine IL-13 production in human mastocytoma cells-1 (Nagarajan et al., 2017). In an allergic asthma model, compound A pre-treatment (10, 20, 30 mg/kg) led to significant reduction of IgE and Th2 cytokines in BAL fluid, level of inflammatory infiltrates and signaling molecules such as ERK1/2, Akt, SAPK/JNK and NF- κ B in lung tissue (Nagarajan and Thangam, 2020).

ZPL-3893787, formerly PF-3893787, is a selective oral H₄-antihistamine, was used daily (30 mg) in adult patients with moderate-to-severe atopic dermatitis in a randomized, double-blind, placebo-controlled, parallel-group study. ZPL-3893787 was well tolerated and showed significant improvement in atopic dermatitis scores (EASI, IGA, SCORAD) compared with placebo group (NCT02424253) (Werfel et al., 2019). Other RCTs in atopic dermatitis patients are ongoing (NCT03517566, NCT03948334).

Effects of several structurally related H₄R ligands of the aminopyrimidine class were evaluated on croton oil-induced ear edema and pruritus in mice. ST-994 and ST-1012 (30 and 100 mg/kg) significantly reduced ear edema. Additionally, ST-994 decreased the inflammatory severity score and in the number of eosinophils in inflamed ear tissue (Adami et al., 2018).

INCB38579 is a pyrimidine-based small molecule antihistamine of the human and rodent H₄R. INCB38579 has good pharmacokinetic properties in rats and mice and showed 80-fold selectivity over the human H₁-H₃R. In *in vitro* experiments, INCB38579 had similar potencies as JNJ7777120 inhibiting histamine-evoked migration of human eosinophils and human and mouse dendritic cells, as well as the shape change of human eosinophils. Furthermore, INCB38579 100 mg/kg were given orally to

CD-1 mice before injection of histamine. INCB38579 and JNJ7777120 significantly reduced the number of scratching bouts. In addition, INCB38579 and JNJ7777120 were able to inhibit carrageenan-induced mechanical hyperalgesia in rats and formalin-induced pain in rats and mice (Shin et al., 2012).

Neumann et al. analyzed the effects of subcutaneous injection of JNJ7777120 in comparison to that of the H₃/H₄-antihistamine thioperamide in a murine asthma model. Thioperamide showed better pharmacokinetics, higher t_{1/2} values and maximal concentration in lung tissue, but weaker beneficial effects as compared to JNJ7777120. Thioperamide reduced only eosinophilia in bronchoalveolar lavage fluid, while JNJ7777120 was in addition able to decrease serum levels of allergen-specific IgE and inflammatory infiltrations in lung tissue (Neumann et al., 2013).

Efficacy and safety of toreforant (JNJ38518168), an oral selective H₄-antihistamine, was evaluated in 162 patients with uncontrolled, eosinophilic asthma in a phase 2a multicenter randomized controlled trial (NCT01823016). Toreforant was well tolerated but failed to provide therapeutic benefit in this population of patients (Kollmeier et al., 2018).

A940894 is another selective H₄R antagonist that binds to both human and rat H₄R. A940894 showed good half-life and oral bioavailability in rats and mice. *In vitro*, it potently blocked histamine-induced calcium mobilization and shape change of mouse bone marrow-derived mast cells and chemotaxis of human eosinophils. In a mouse mast cell-dependent model of zymosan-induced peritonitis, A940894 markedly suppressed neutrophil influx and decreased intraperitoneal prostaglandin D₂ levels (Strakhova et al., 2009).

Effects of VUF6002 (JNJ10191584) were evaluated in the rat model of carrageenan-induced inflammation and thermal hyperalgesia. VUF6002 (10 mg/kg, subcutaneously as well as JNJ7777120 (10 and 30 mg/kg, subcutaneously) were able to significantly reduce paw edema and hyperalgesia (Coruzzi et al., 2007).

UR-63325, a selective H₄-antihistamine, inhibited degranulation of bone marrow-derived mast cells (Mirzahosseini et al., 2013) and histamine-induced eosinophil chemotaxis *in vitro*, and 4-methyl-histamine-induced pruritus in rat (Salcedo et al., 2013). It is also demonstrated efficacy in several asthma models in rodents (Salcedo et al., 2013). In healthy volunteers, UR-63325 showed a linear pharmacokinetic profile suitable for once daily dosing, good safety, and pharmacodynamic results indicating that full H₄R blockage was achieved with single doses up to 100 mg. (Salcedo et al., 2013) Efficacy of UR-63325 was evaluated in a RCT, which included patients with allergic rhinitis, but no results have been published (NCT01260753).

Orally administered compound 48 exhibited strong antipruritic and anti-inflammation activity in several mouse models of atopic dermatitis (Ko et al., 2018).

Several other potent H₄-antihistamines have been reported including A987306 (Liu et al., 2008), A943931 (Zhang et al., 2019), SENS-111 (Petremann et al., 2020) and TR-7 (Popiolek-Barczyk et al., 2018).

5.28.6.4 H₄-Antihistamines in other diseases

Apart from allergic diseases, some H₄-antihistamines showed beneficial effects in other disorders. For example, in a mice model of autism, JNJ7777120 treatment with irradiation exposure increased social interactions in BTBR mice blocking inflammatory cytokine production and transcription factor signaling compared to that in irradiation-exposed BTBR mice (Ahmad et al., 2019). JNJ7777120 was effective in cisplatin-induced anorexia (Yamamoto et al., 2019) and had protective properties in rat models of Parkinson-like pathology, transient cerebral ischemia and age-related macular degeneration (Zhou et al., 2019; Kaneko et al., 2014; Dettori et al., 2018). JNJ39758979 might be a promising medication for diabetic nephropathy (Pini et al., 2018). Toreforant has been administered with limited efficacy in patients with rheumatoid arthritis and psoriasis (Frankel et al., 2018; Boyle et al., 2019). SENS-111, a novel selective oral H₄-antihistamine, improved vertigo symptoms in a translational *in vivo* model of unilateral vestibular loss (Petremann et al., 2020; Venail et al., 2018).

5.28.7 Conclusion

In conclusion, H₁, H₂ and H₃-antihistamines are well established in widespread clinical diseases. In contrast H₄-antihistamines are in the dawn of their development but have a very promising future.

Disclosure of Potential Conflict of Interest

Martin Church: speaker or consultant for Almirall, FAES Pharma, Menarini, Moxie, MSD, Novartis, UCB Pharma, Sanofi-Aventis and Uriach.

Stefan Frischbutter: no conflicts of interest regarding any aspects of this study.

Pavel Kolkhir: personal fees from Novartis and Roche, outside the submitted work.

Marcus Maurer: grants and/or personal fees from Allakos, Amgen, Aralez, AstraZeneca, Celldex, CSL Behring, FAES, Genentech, GInnovation, Innate Pharma, Menarini, Leo Pharma, Lilly, Moxie, MSD, Roche, Sanofi, Third Harmonic Bio, UCB, Uriach.

See Also: 5.29. Chronic Urticaria

References

- Adam, K., Oswald, I., 1986. The hypnotic effects of an antihistamine: Promethazine. *British Journal of Clinical Pharmacology* 22, 715–717.
- Adami, M., Micheloni, C., Grandi, D., Stark, H., 2018. Differential effects of functionally different histamine H4 receptor ligands on acute irritant dermatitis in mice. *Naunyn-Schmiedeberg's Archives of Pharmacology* 391, 1387–1397.
- Ahmad, S.F., Nadeem, A., Ansari, M.A., Bakheet, S.A., Al-Mazroua, H.A., Khan, M.R., Alasmari, A.F., Alanazi, W.A., As Sobeai, H.M., Attia, S.M., 2019. The histamine-4 receptor antagonist JNJ777120 prevents immune abnormalities by inhibiting ROR γ T/T-bet transcription factor signaling pathways in BTBR T+ ltr3tf/J mice exposed to gamma rays. *Molecular Immunology* 114, 561–570.
- Akamine, Y., Miura, M., Komori, H., Tamai, I., Ieiri, I., Yasui-Furukori, N., Uno, T., 2015. The change of pharmacokinetics of fexofenadine enantiomers through the single and simultaneous grapefruit juice ingestion. *Drug Metabolism and Pharmacokinetics* 30, 352–357.
- Akdis, C.A., Simons, F.E., 2006. Histamine receptors are hot in immunopharmacology. *European Journal of Pharmacology* 533, 69–76.
- Arrang, J.M., Garbarg, M., Schwartz, J.C., 1983. Auto-inhibition of brain histamine release mediated by a novel class (H3) of histamine receptor. *Nature* 302, 832–837.
- Ash, A.S., Schild, H.O., 1966. Receptors mediating some actions of histamine. *British Journal of Pharmacology and Chemotherapy* 27, 427–439.
- Asher, M.I., Montefort, S., Bjorksten, B., Lai, C.K., Strachan, D.P., Weiland, S.K., Williams, H., ISAAC Phase Three Study Group, 2006. Worldwide time trends in the prevalence of symptoms of asthma, allergic rhinoconjunctivitis, and eczema in childhood: ISAAC phases one and three repeat multicountry cross-sectional surveys. *Lancet* 368, 733–743.
- Bachert, C., 2009. A review of the efficacy of desloratadine, fexofenadine, and levocetirizine in the treatment of nasal congestion in patients with allergic rhinitis. *Clinical Therapeutics* 31, 921–944.
- Bachert, C., Maurer, M., 2010. Safety and efficacy of desloratadine in subjects with seasonal allergic rhinitis or chronic urticaria: Results of four postmarketing surveillance studies. *Clinical Drug Investigation* 30, 109–122.
- Bachert, C., Bousquet, J., Canonica, G.W., Durham, S.R., Klimek, L., Mullol, J., Van Cauwenberge, P.B., Van Hamme, G., XPERT Study Group, 2014. Levocetirizine improves quality of life and reduces costs in long-term management of persistent allergic rhinitis. *The Journal of Allergy and Clinical Immunology* 114, 838–844.
- Baena-Cagnani, C.E., Berger, W.E., DuBuske, L.M., Gurne, S.E., Strzysak, P., Lorber, R., Danzig, M., 2003. Comparative effects of desloratadine versus montelukast on asthma symptoms and use of beta 2-agonists in patients with seasonal allergic rhinitis and asthma. *International Archives of Allergy and Immunology* 130, 307–313.
- Bakker, R.A., Schoonus, S.B., Smit, M.J., Timmerman, H., Leurs, R., 2001. Histamine H(1)-receptor activation of nuclear factor-kappa B: Roles for G beta gamma- and G alpha(q/11)-subunits in constitutive and agonist-mediated signaling. *Molecular Pharmacology* 60, 1133–1142.
- Banu, Y., Watanabe, T., 1999. Augmentation of antigen receptor-mediated responses by histamine H1 receptor signaling. *The Journal of Experimental Medicine* 189, 673–682.
- Barchuk, W.T., Salapatek, A.M., Ge, T., D'Angelo, P., Liu, X., 2013. A proof-of-concept study of the effect of a novel H3-receptor antagonist in allergen-induced nasal congestion. *The Journal of Allergy and Clinical Immunology* 132 (838-846), e1–e6.
- Berger, W.E., Lumry, W.R., Meltzer, E.O., Pearlman, D.S., 2006. Efficacy of desloratadine, 5 mg, compared with fexofenadine, 180 mg, in patients with symptomatic seasonal allergic rhinitis. *Allergy and Asthma Proceedings* 27, 214–223.
- Black, J.W., Duncan, W.A., Durant, C.J., Ganellin, C.R., Parsons, E.M., 1972. Definition and antagonism of histamine H 2-receptors. *Nature* 236, 385–390.
- Bojkowski, C.J., Gibbs, T.G., Hellstern, K.H., Major, E.W., Mullinger, B., 1989. Acrivastine in allergic rhinitis: A review of clinical experience. *The Journal of International Medical Research* 17 (Supplement 2), 54B–68B.
- Bosma, R., Van Den Bor, J., Vischer, H.F., Labeaga, L., Leurs, R., 2018. The long duration of action of the second generation antihistamine bilastine coincides with its long residence time at the histamine H1 receptor. *European Journal of Pharmacology* 838, 107–111.
- Bousquet, J., Khaltaev, N., Cruz, A.A., Denburg, J., Fokkens, W.J., Togias, A., Zuberbier, T., Baena-Cagnani, C.E., Canonica, G.W., Van Weel, C., Agache, I., Ait-Khaled, N., Bachert, C., Blaiss, M.S., Bonini, S., Boulet, L.P., Bousquet, P.J., Camargos, P., Carlsen, K.H., Chen, Y., Custovic, A., Dahl, R., Demoly, P., Douagui, H., Durham, S.R., Van Wijk, R.G., Kalayci, O., Kaliner, M.A., Kim, Y.Y., Kowalski, M.L., Kuna, P., Le, L.T., Lemiere, C., Li, J., Lockey, R.F., Mavale-Manuel, S., Meltzer, E.O., Mohammad, Y., Mullol, J., Naclerio, R., O'Hehir, R.E., Ohta, K., Ouedraogo, S., Palkonen, S., Papadopoulos, N., Passalacqua, G., Pawankar, R., Popov, T.A., Rabe, K.F., Rosado-Pinto, J., Scadding, G.K., Simons, F.E., Toskala, E., Valovirta, E., Van Cauwenberge, P., Wang, D.Y., Wickman, M., Yawn, B.P., Yorgancioglu, A., Yusuf, O.M., Zar, H., Annesi-Maesano, I., Bateman, E.D., Ben Kheder, A., Boakye, D.A., Bouchard, J., Burney, P., Busse, W.W., Chan-Yeung, M., Chavannes, N.H., Chuchalin, A., Dolen, W.K., Emuzyte, R., Grouse, L., Humbert, M., Jackson, C., Johnston, S.L., Keith, P.K., Kemp, J.P., Klossek, J.M., Larenas-Linnemann, D., Lipworth, B., Malo, J.L., Marshall, G.D., Nasipit, C., Nekam, K., Niggemann, B., Nizankowska-Mogilnicka, E., Okamoto, Y., Orr, M.P., Potter, P., Price, D., Stoloff, S.W., Vandenplas, O., Viegi, G., Williams, D., World Health, O., Galen & Allergen, 2008. Allergic Rhinitis and its Impact on Asthma (ARIA) 2008 update (in collaboration with the World Health Organization, GA(2)LEN and AllerGen). *Allergy* 63 (Supplement 86), 8–160.
- Bovet, D., 1950. Introduction to antihistamine agents and antergan derivative. *Annals of the New York Academy of Sciences* 50, 1089–1126.
- Boyle, J., Eriksson, M., Stanley, N., Fujita, T., Kumagi, Y., 2006. Allergy medication in Japanese volunteers: Treatment effect of single doses on nocturnal sleep architecture and next day residual effects. *Current Medical Research and Opinion* 22, 1343–1351.
- Boyle, D.L., Deprimo, S.E., Calderon, C., Chen, D., Dunford, P.J., Barchuk, W., Firestein, G.S., Thurmond, R.L., 2019. Toreforant, an orally active histamine H4-receptor antagonist, in patients with active rheumatoid arthritis despite methotrexate: Mechanism of action results from a phase 2, multicenter, randomized, double-blind, placebo-controlled synovial biopsy study. *Inflammation Research* 68, 261–274.
- Brimblecombe, R.W., Duncan, W.A., Durant, G.J., Ganellin, C.R., Parsons, M.E., Black, J.W., 1975. The pharmacology of cimetidine, a new histamine H2-receptor antagonist. *British Journal of Pharmacology* 53, 435P–436P.
- Brioni, J.D., Esbenschade, T.A., Garrison, T.R., Bitner, S.R., Cowart, M.D., 2011. Discovery of histamine H3 antagonists for the treatment of cognitive disorders and Alzheimer's disease. *The Journal of Pharmacology and Experimental Therapeutics* 336, 38–46.
- Brogden, R.N., McTavish, D., 1991. Acrivastine. A review of its pharmacological properties and therapeutic efficacy in allergic rhinitis, urticaria and related disorders. *Drugs* 41, 927–940.
- Broide, D., 1995. Clinical studies with cetirizine in allergic rhinitis and chronic urticaria. *Allergy* 50, 31–35.
- Brown, R.E., Stevens, D.R., Haas, H.L., 2001. The physiology of brain histamine. *Progress in Neurobiology* 63, 637–672.
- Brozek, J.L., Bousquet, J., Agache, I., Agarwal, A., Bachert, C., Bosnic-Anticevich, S., Brignardello-Petersen, R., Canonica, G.W., Casale, T., Chavannes, N.H., Correia De Sousa, J., Cruz, A.A., Cuello-Garcia, C.A., Demoly, P., Dykewicz, M., Etzeandia-Ikobaltzeta, I., Florez, I.D., Fokkens, W., Fonseca, J., Hellings, P.W., Klimek, L., Kowalski, S., Kuna, P., Laisaar, K.T., Larenas-Linnemann, D.E., Lodrup Carlsen, K.C., Manning, P.J., Meltzer, E., Mullol, J., Muraro, A., O'Hehir, R., Ohta, K., Panzner, P., Papadopoulos, N., Park, H.S., Passalacqua, G., Pawankar, R., Price, D., Riva, J.J., Roldan, Y., Ryan, D., Sadeghirad, B., Samolinski, B., Schmid-Grendelmeier, P., Sheikh, A., Togias, A., Valero, A., Valiulis, A., Valovirta, E., Ventresca, M., Wallace, D., Wasserman, S., Wickman, M., Wiercioch, W., Yepes-Nunez, J.J., Zhang, L., Zhang, Y., Zidarn, M., Zuberbier, T., Schunemann, H.J., 2017. Allergic rhinitis and its impact on asthma (ARIA) guidelines-2016 revision. *The Journal of Allergy and Clinical Immunology* 140, 950–958.
- Bryce, P.J., Mathias, C.B., Harrison, K.L., Watanabe, T., Geha, R.S., Oettgen, H.C., 2006. The H1 histamine receptor regulates allergic lung responses. *The Journal of Clinical Investigation* 116, 1624–1632.
- Burton, P.S., Crean, C., Kagey, M., Neilsen, J.W., 2007. Permeability characteristics of bilastine, a potent and selective H1 receptor antagonist, in caco-2 cells. *The AAPS Journal* 9 (Supplement 2), 2910.
- Canonica, G.W., Fumagalli, F., Guerra, L., Baiardini, I., Compalati, E., Rogkakou, A., Massacane, P., Gamalero, C., Riccio, A.M., Scordamaglia, A., Passalacqua, G., Global Allergy and Asthma European Network, 2008. Levocetirizine in persistent allergic rhinitis: Continuous or on-demand use? A pilot study. *Current Medical Research and Opinion* 24, 2829–2839.
- Carpenter, G.B., Bunker-Soler, A.L., Nelson, H.S., 1983. Evaluation of combined H1- and H2-receptor blocking agents in the treatment of seasonal allergic rhinitis. *The Journal of Allergy and Clinical Immunology* 71, 412–417.

- Cataldi, M., Maurer, M., Tagliatela, M., Church, M.K., 2019. Cardiac safety of second-generation H1-antihistamines when up-dosed in chronic spontaneous urticaria. *Clinical and Experimental Allergy* 49, 1615–1623.
- Chen, C., 2008. Physicochemical, pharmacological and pharmacokinetic properties of the zwitterionic antihistamines cetirizine and levocetirizine. *Current Medicinal Chemistry* 15, 2173–2191.
- Chen, C., Hanson, E., Watson, J.W., Lee, J.S., 2003. P-glycoprotein limits the brain penetration of non-sedating but not sedating H1-antagonists. *Drug Metabolism and Disposition* 31, 312–318.
- Church, M.K., 1999. Non-H1-receptor effects of antihistamines. *Clinical and Experimental Allergy* 29 (Supplement 3), 39–48.
- Church, M.K., 2011. Safety and efficacy of bilastine: A new H(1)-antihistamine for the treatment of allergic rhinoconjunctivitis and urticaria. *Expert Opinion on Drug Safety* 10, 779–793.
- Church, M.K., Labeaga, L., 2017. Bilastine: A new H1-antihistamine with an optimal profile for up-dosing in urticaria. *Journal of the European Academy of Dermatology and Venereology* 31, 1447–1452.
- Church, M.K., Maurer, M., 2012. H(1)-antihistamines and urticaria: How can we predict the best drug for our patient? *Clinical and Experimental Allergy* 42, 1423–1429.
- Church, M.K., Bewley, A.P., Clough, G.F., Burrows, L.J., Ferdinand, S.I., Petersen, L.J., 1997. Studies into the mechanisms of dermal inflammation using cutaneous microdialysis. *International Archives of Allergy and Immunology* 113, 131–133.
- Church, M.K., Maurer, M., Simons, F.E., Bindslev-Jensen, C., Van Cauwenberge, P., Bousquet, J., Holgate, S.T., Zuberbier, T., Global Allergy and Asthma European Network, 2010. Risk of first-generation H(1)-antihistamines: A GA(2)LEN position paper. *Allergy* 65, 459–466.
- Church, M.K., Weller, K., Stock, P., Maurer, M., 2011. Chronic spontaneous urticaria in children: Itching for insight. *Pediatric Allergy and Immunology* 22, 1–8.
- Church, M.K., Kolkhir, P., Metz, M., Maurer, M., 2018. The role and relevance of mast cells in urticaria. *Immunological Reviews* 282, 232–247.
- Corcostegui, R., Labeaga, L., Innerarity, A., Berisa, A., Orjales, A., 2005. Preclinical pharmacology of bilastine, a new selective histamine H1 receptor antagonist: Receptor selectivity and in vitro antihistaminic activity. *Drugs in R&D* 6, 371–384.
- Coruzzi, G., Adami, M., Guaita, E., De Esch, I.J.P., Leurs, R., 2007. Anti-inflammatory and antinociceptive effects of the selective histamine H4-receptor antagonists JNJ7777120 and VUF6002 in a rat model of carrageenan-induced acute inflammation. *European Journal of Pharmacology* 563, 240–244.
- Cowden, J.M., Zhang, M., Dunford, P.J., Thurmond, R.L., 2010. The histamine H4 receptor mediates inflammation and pruritus in Th2-dependent dermal inflammation. *The Journal of Investigative Dermatology* 130, 1023–1033.
- Crean, C., Valiente, R., Sologuren, A., Mclaverty, D., 2007. Effect of grapefruit juice on the pharmacokinetics of bilastine. *Journal of Clinical Pharmacology* 47, 1198.
- Crowe, A., Wright, C., 2012. The impact of P-glycoprotein mediated efflux on absorption of 11 sedating and less-sedating antihistamines using Caco-2 monolayers. *Xenobiotica* 42, 538–549.
- Curran, M.P., Scott, L.J., Perry, C.M., 2004. Cetirizine: A review of its use in allergic disorders. *Drugs* 64, 523–561.
- Cvetkovic, M., Leake, B., Fromm, M.F., Wilkinson, G.R., Kim, R.B., 1999. OATP and P-glycoprotein transporters mediate the cellular uptake and excretion of fexofenadine. *Drug Metabolism and Disposition* 27, 866–871.
- Denham, K.J., Boutsouki, P., Clough, G.F., Church, M.K., 2003. Comparison of the effects of desloratadine and levocetirizine on histamine-induced wheal, flare and itch in human skin. *Inflammation Research* 52, 424–427.
- Dettoni, I., Gaviano, L., Melani, A., Lucarini, L., Durante, M., Masini, E., Pedata, F., 2018. A selective histamine H4 receptor antagonist, JNJ7777120, is protective in a rat model of transient cerebral ischemia. *Frontiers in Pharmacology* 9, 1231.
- Devillier, P., Roche, N., Faisy, C., 2008. Clinical pharmacokinetics and pharmacodynamics of desloratadine, fexofenadine and levocetirizine: A comparative review. *Clinical Pharmacokinetics* 47, 217–230.
- Dimitriadou, V., Rouleau, A., Dam Trung Tuong, M., Newlands, G.J., Miller, H.R., Luffau, G., Schwartz, J.C., Garbag, M., 1994. Functional relationship between mast cells and C-sensitive nerve fibres evidenced by histamine H3-receptor modulation in rat lung and spleen. *Clinical Science (London, England)* 87, 151–163.
- Dresser, G.K., Kim, R.B., Bailey, D.G., 2005. Effect of grapefruit juice volume on the reduction of fexofenadine bioavailability: Possible role of organic anion transporting polypeptides. *Clinical Pharmacology and Therapeutics* 77, 170–177.
- Druce, H.M., Thoden, W.R., Mure, P., Furey, S.A., Lockhart, E.A., Xie, T., Galant, S., Prenner, B.M., Weinstein, S., Ziering, R., Brandon, M.L., 1998. Brompheniramine, loratadine, and placebo in allergic rhinitis: A placebo-controlled comparative clinical trial. *Journal of Clinical Pharmacology* 38, 382–389.
- DuBuske, L.M., 1999. Second-generation antihistamines: The risk of ventricular arrhythmias. *Clinical Therapeutics* 21, 281–295.
- Ducharme, E.E., Weinberg, J.M., 2009. Levocetirizine for chronic idiopathic urticaria: A review. *Journal of Drugs in Dermatology* 8, 243–247.
- Dunford, P.J., Williams, K.N., Desai, P.J., Karlsson, L., McQueen, D., Thurmond, R.L., 2007. Histamine H4 receptor antagonists are superior to traditional antihistamines in the attenuation of experimental pruritus. *The Journal of Allergy and Clinical Immunology* 119, 176–183.
- Durante, M., Sgambellone, S., Lanzi, C., Nardini, P., Pini, A., Moroni, F., Masini, E., Lucarini, L., 2019. Effects of PARP-1 deficiency and histamine H4 receptor inhibition in an inflammatory model of lung fibrosis in mice. *Frontiers in Pharmacology* 10, 525.
- Ebenezer, A.J., Arunachalam, P., Elden, B.T., 2017. H4R activation utilizes distinct signaling pathways for the production of RANTES and IL-13 in human mast cells. *Journal of Receptor and Signal Transduction Research* 37, 133–140.
- Ebenezer, A.J., Prasad, K., Rajan, S., Thangam, E.B., 2018. Silencing of H4R inhibits the production of IL-1 β through SAPK/JNK signaling in human mast cells. *Journal of Receptor and Signal Transduction Research* 38, 204–212.
- Ehling, S., Rossbac, K., Dunston, S.M., Stark, H., Bäumer, W., 2016. Allergic inflammation is augmented via histamine H4 receptor activation: The role of natural killer cells in vitro and in vivo. *Journal of Dermatological Science* 83, 106–115.
- Emanuel, M.B., 1999. Histamine and the antiallergic antihistamines: A history of their discoveries. *Clinical and Experimental Allergy* 29 (Supplement 3), 1–11.
- Farre, M., Perez-Mana, C., Papaseit, E., Menoyo, E., Perez, M., Martin, S., Bullich, S., Rojas, S., Herance, J.R., Trampal, C., Labeaga, L., Valiente, R., 2014. Bilastine vs. hydroxyzine: Occupation of brain histamine H1-receptors evaluated by positron emission tomography in healthy volunteers. *British Journal of Clinical Pharmacology* 78, 970–980.
- Fedorowicz, Z., Van Zuuren, E.J., Hu, N., 2012. Histamine H2-receptor antagonists for urticaria. *Cochrane Database of Systematic Reviews*. CD008596.
- Frankel, E., Song, M., Li, S., Jiang, J., Thurmond, R.L., Randazzo, B., 2018. Efficacy and safety of toreforant, a selective histamine H4 receptor antagonist, for the treatment of moderate-to-severe plaque psoriasis: Results from a phase 2 multicenter, randomized, double-blind, placebo-controlled trial. *Journal of Drugs in Dermatology* 17, 873–879.
- Geha, R.S., Meltzer, E.O., 2001. Desloratadine: A new, non-sedating, oral antihistamine. *The Journal of Allergy and Clinical Immunology* 107, 751–762.
- Gibson, J.R., Manna, V.K., Salisbury, J., 1989. Acrivastine: A review of its dermatopharmacology and clinical activity. *The Journal of International Medical Research* 17 (Supplement 2), 28B–34B.
- Gillard, M., Chatelain, P., 2006. Changes in pH differently affect the binding properties of histamine H1 receptor antagonists. *European Journal of Pharmacology* 530, 205–214.
- Gillard, M., Van Der Perren, C., Moguilevsky, N., Massingham, R., Chatelain, P., 2002. Binding characteristics of cetirizine and levocetirizine to human H(1) histamine receptors: Contribution of Lys(191) and Thr(194). *Molecular Pharmacology* 61, 391–399.
- Gillard, M., Christophe, B., Wels, B., Peck, M., Massingham, R., Chatelain, P., 2003. H1 antagonists: Receptor affinity versus selectivity. *Inflammation Research* 52 (Supplement 1), S49–S50.
- Gillard, M., Benedetti, M.S., Chatelain, P., Baltes, E., 2005. Histamine H1 receptor occupancy and pharmacodynamics of second generation H1-antihistamines. *Inflammation Research* 54, 367–369.
- Gillman, S., Gillard, M., Strolin Benedetti, M., 2009. The concept of receptor occupancy to predict clinical efficacy: A comparison of second generation H1 antihistamines. *Allergy and Asthma Proceedings* 30, 366–376.

- Giustizieri, M.L., Albanesi, C., Fluhr, J., Gisondi, P., Norgauer, J., Girolomoni, G., 2004. H1 histamine receptor mediates inflammatory responses in human keratinocytes. *The Journal of Allergy and Clinical Immunology* 114, 1176–1182.
- Glatzer, F., Gschwandtner, M., Ehling, S., Rossbach, K., Janik, K., Klos, A., Bäumer, W., Kietzmann, M., Werfel, T., Gutzmer, R., 2013. Histamine induces proliferation in keratinocytes from patients with atopic dermatitis through the histamine 4 receptor. *The Journal of Allergy and Clinical Immunology* 132, 1358–1367.
- Gonzalez-Nunez, V., Bachert, C., Mullol, J., 2016. Rupatadine: Global safety evaluation in allergic rhinitis and urticaria. *Expert Opinion on Drug Safety* 15, 1439–1448.
- Grant, J.A., Riethuisen, J.M., Moulart, B., Devos, C., 2002. A double-blind, randomized, single-dose, crossover comparison of levocetirizine with ebastine, fexofenadine, loratadine, mizolastine, and placebo: Suppression of histamine-induced wheal-and-flare response during 24 hours in healthy male subjects. *Annals of Allergy, Asthma & Immunology* 88, 190–197.
- Gray, S.L., Anderson, M.L., Dublin, S., Hanlon, J.T., Hubbard, R., Walker, R., Yu, O., Crane, P.K., Larson, E.B., 2015. Cumulative use of strong anticholinergics and incident dementia: A prospective cohort study. *JAMA Internal Medicine* 175, 401–407.
- Grosicki, M., Wójcik, T., Chlopicki, S., Kieć-Kononowicz, K., 2016. In vitro study of histamine and histamine receptor ligands influence on the adhesion of purified human eosinophils to endothelium. *European Journal of Pharmacology* 777, 49–59.
- Gutzmer, R., Mommert, S., Gschwandtner, M., Zwingmann, K., Stark, H., Werfel, T., 2009. The histamine H4 receptor is functionally expressed on T(H)2 cells. *The Journal of Allergy and Clinical Immunology* 123, 619–625.
- Haas, H., Panula, P., 2003. The role of histamine and the tuberomammillary nucleus in the nervous system. *Nature Reviews. Neuroscience* 4, 121–130.
- Halpern, B.N., 1942. Les antihistaminiques de synthèse. Essais de chimiothérapie des états allergiques. *Archives Internationales de Pharmacodynamie et de Thérapie* 681, 339–408.
- Hiraoka, K., Tashiro, M., Grobosch, T., Maurer, M., Oda, K., Toyohara, J., Ishii, K., Ishiwata, K., Yanai, K., 2015. Brain histamine H1 receptor occupancy measured by PET after oral administration of levocetirizine, a non-sedating antihistamine. *Expert Opinion on Drug Safety* 14, 199–206.
- Hofstra, C.L., Desai, P.J., Thurmond, R.L., Fung-Leung, W.-P., 2003. Histamine H4 receptor mediates chemotaxis and calcium mobilization of mast cells. *The Journal of Pharmacology and Experimental Therapeutics* 305, 1212–1221.
- Holgate, S.T., Canonica, G.W., Simons, F.E., Tagliabue, M., Tharp, M., Timmerman, H., Yanai, K., Consensus Group on New-Generation Antihistamines, 2003. Consensus Group on New-Generation Antihistamines (CONGA): Present status and recommendations. *Clinical and Experimental Allergy* 33, 1305–1324.
- Hong, H.K., Jo, S.H., 2009. Block of HERG K channel by classic histamine h(1) receptor antagonist chlorpheniramine. *Korean Journal of Physiology & Pharmacology* 13, 215–220.
- Horio, S., Fujimoto, K., Mizuguchi, H., Fukui, H., 2010. Interleukin-4 up-regulates histamine H1 receptors by activation of H1 receptor gene transcription. *Naunyn-Schmiedeberg's Archives of Pharmacology* 381, 305–313.
- Huang, K., Hu, D.-D., Bai, D., Wu, Z.-Y., Chen, Y.-Y., Zhang, Y.-J., Lv, X., Wang, Q.-X., Zhang, L., 2018. Persistent extracellular signal-regulated kinase activation by the histamine H4 receptor in spinal neurons underlies chronic itch. *The Journal of Investigative Dermatology* 138, 1843–1850.
- Hulhoven, R., Rosillon, D., Letiexhe, M., Meeus, M.A., Daoust, A., Stockis, A., 2007. Levocetirizine does not prolong the QT/QTc interval in healthy subjects: Results from a thorough QT study. *European Journal of Clinical Pharmacology* 63, 1011–1017.
- Hunt, J.F., Fang, K., Malik, R., Snyder, A., Malhotra, N., Platts-Mills, T.A., Gaston, B., 2000. Endogenous airway acidification. Implications for asthma pathophysiology. *American Journal of Respiratory and Critical Care Medicine* 161, 694–699.
- Izquierdo, I., Merlos, M., Garcia-Rafanell, J., 2003. Rupatadine: A new selective histamine H1 receptor and platelet-activating factor (PAF) antagonist. A review of pharmacological profile and clinical management of allergic rhinitis. *Drugs Today* 39, 451–468.
- Jauregizar, N., De La Fuente, L., Lucero, M.L., Sologuren, A., Leal, N., Rodriguez, M., 2009. Pharmacokinetic-pharmacodynamic modelling of the antihistaminic (H1) effect of bilastine. *Clinical Pharmacokinetics* 48, 543–554.
- Jauregui, I., Ferrer, M., Bartra, J., Del Cuvillo, A., Davila, I., Montoro, J., Mullol, J., Sastre, J., Valero, A., 2013. Bilastine for the treatment of urticaria. *Expert Opinion on Pharmacotherapy* 14, 1537–1544.
- Jemima, E.A., Prema, A., Thangam, E.B., 2014. Functional characterization of histamine H4 receptor on human mast cells. *Molecular Immunology* 62, 19–28.
- Jian, T., Yang, N., Yang, Y., Zhu, C., Yuan, X., Yu, G., Wang, C., Wang, Z., Shi, H., Tang, M., He, Q., Lan, L., Wu, G., Tang, Z., 2015. TRPV1 and PLC participate in histamine H4 receptor-induced itch. *Neural Plasticity* 2016, 2016.
- Jo, S.H., Hong, H.K., Chong, S.H., Lee, H.S., Choe, H., 2009. H(1) antihistamine drug promethazine directly blocks hERG K(+) channel. *Pharmacological Research* 60, 429–437.
- Juniper, E.F., Stahl, E., Doty, R.L., Simons, F.E., Allen, D.B., Howarth, P.H., 2005. Clinical outcomes and adverse effect monitoring in allergic rhinitis. *The Journal of Allergy and Clinical Immunology* 115, S390–S413.
- Jutel, M., Klunker, S., Akdis, M., Malolepszy, J., Thomet, O.A., Zak-Nejmark, T., Blaser, K., Akdis, C.A., 2001. Histamine upregulates Th1 and downregulates Th2 responses due to different patterns of surface histamine 1 and 2 receptor expression. *International Archives of Allergy and Immunology* 124, 190–192.
- Kanda, N., Watanabe, S., 2003. Histamine enhances the production of nerve growth factor in human keratinocytes. *The Journal of Investigative Dermatology* 121, 570–577.
- Kaneko, H., Ye, F., Iijima, R., Kachi, S., Kato, S., Nagaya, M., Higuchi, A., Terasaki, H., 2014. Histamine H4 receptor as a new therapeutic target for choroidal neovascularization in age-related macular degeneration. *British Journal of Pharmacology* 171, 3754–3763.
- Kapp, A., Wedi, B., 2004. Chronic urticaria: Clinical aspects and focus on a new antihistamine, levocetirizine. *Journal of Drugs in Dermatology* 3, 632–639.
- Katelaris, C.H., Lee, B.W., Potter, P.C., Maspero, J.F., Cingi, C., Lopatin, A., Safer, M., Xu, G., Walters, R.D., 2012. Prevalence and diversity of allergic rhinitis in regions of the world beyond Europe and North America. *Clinical and Experimental Allergy* 42, 186–207.
- Kawashima, M., Harada, S., Tango, T., 2002. Review of fexofenadine in the treatment of chronic idiopathic urticaria. *International Journal of Dermatology* 41, 701–706.
- Kay, G.G., Berman, B., Mockoviak, S.H., Morris, C.E., Reeves, D., Starbuck, V., Sukenik, E., Harris, A.G., 1997. Initial and steady-state effects of diphenhydramine and loratadine on sedation, cognition, mood, and psychomotor performance. *Archives of Internal Medicine* 157, 2350–2356.
- Kay, L.J., Suvarna, S.K., Peachell, P.T., 2018. Histamine H4 receptor mediates chemotaxis of human lung mast cells. *European Journal of Pharmacology* 837, 38–44.
- Ko, K., Kim, H.-J., Ho, P.-S., Lee, S.O., Lee, J.-E., Min, C.-R., Kim, Y.C., Yoon, J.-H., Park, E.-J., Kwon, Y.-J., Yun, J.-H., Yoon, D.-O., Kim, J.-S., Park, W.-S., Oh, S.-S., Song, Y.-M., Cho, W.-K., Morikawa, K., Lee, K.-J., Park, C.-H., 2018. Discovery of a novel highly selective histamine H4 receptor antagonist for the treatment of atopic dermatitis. *Journal of Medicinal Chemistry* 61, 2949–2961.
- Köchling, H., Schaper, K., Wilzopolski, J., Gutzmer, R., Werfel, T., Bäumer, W., Kietzmann, M., Rossbach, K., 2017. Combined treatment with H1 and H4 receptor antagonists reduces inflammation in a mouse model of atopic dermatitis. *Journal of Dermatological Science* 87, 130–137.
- Kollmeier, A., Francke, K., Chen, B., Dunford, P.J., Greenspan, A.J., Xia, Y., Xu, X.L., Zhou, B., Thurmond, R.L., 2014. The histamine H4 receptor antagonist, JNJ 39758979, is effective in reducing histamine-induced pruritus in a randomized clinical study in healthy subjects. *The Journal of Pharmacology and Experimental Therapeutics* 350, 181–187.
- Kollmeier, A.P., Barnathan, E.S., O'BRIEN, C., Chen, B., Xia, Y.K., Zhou, B., Loza, M.J., Silkoff, P.E., Ge, M., Thurmond, R.L., 2018. A phase 2a study of tofevorant, a histamine H4 receptor antagonist, in eosinophilic asthma. *Annals of Allergy, Asthma & Immunology* 121, 568–574.
- Kovacova-Hanusikova, E., Gavliakova, S., Buday, T., Kosutova, P., Mokry, D., Urbanova-Fulmekova, A., Mokry, J., Plevkova, J., 2015. The effect of selective antagonist of H4 receptor JNJ7777120 on nasal symptoms, cough, airway reactivity and inflammation in guinea pigs. *Respiratory Physiology & Neurobiology* 216, 9–14.
- Krause, K., Spohr, A., Zuberbier, T., Church, M.K., Maurer, M., 2013. Up-dosing with bilastine results in improved effectiveness in cold contact urticaria. *Allergy* 68, 921–928.
- Lappin, G., Shishikura, Y., Jochemsen, R., Weaver, R.J., Gesson, C., Houston, B., Oosterhuis, B., Bjerrum, O.J., Rowland, M., Garner, C., 2010. Pharmacokinetics of fexofenadine: Evaluation of a microdose and assessment of absolute oral bioavailability. *European Journal of Pharmaceutical Sciences* 40, 125–131.
- Lebrun-Vignes, B., Diquet, B., Chosidow, O., 2001. Clinical pharmacokinetics of mizolastine. *Clinical Pharmacokinetics* 40, 501–507.
- Leurs, R., Church, M.K., Tagliabue, M., 2002. H1-antihistamines: Inverse agonism, anti-inflammatory actions and cardiac effects. *Clinical and Experimental Allergy* 32, 489–498.

- Leurs, R., Chazot, P.L., Shenton, F.C., Lim, H.D., De Esch, I.J., 2009. Molecular and biochemical pharmacology of the histamine H4 receptor. *British Journal of Pharmacology* 157, 14–23.
- Lichtenstein, L.M., Gillespie, E., 1975. The effects of the H1 and H2 antihistamines on "allergic" histamine release and its inhibition by histamine. *The Journal of Pharmacology and Experimental Therapeutics* 192, 441–450.
- Lieberman, P., 2011. The basics of histamine biology. *Annals of Allergy, Asthma & Immunology* 106, S2–S5.
- Limon, L., Kockler, D.R., 2003. Desloratadine: A non-sedating antihistamine. *The Annals of Pharmacotherapy* 37, 237–246 quiz 313–6.
- Lippert, U., Artuc, M., Grützkau, A., Babina, M., Guhl, S., Haase, I., Blaschke, V., Zachmann, K., Knosalla, M., Middel, P., Krüger-Krasagakis, S., Henz, B.M., 2004. Human skin mast cells express H2 and H4, but not H3 receptors. *The Journal of Investigative Dermatology* 123, 116–123.
- Liu, H., Altenbach, R.J., Carr, T.L., Chandran, P., Hsieh, G.C., Lewis, L.G.R., Manelli, A.M., Millicic, I., Marsh, K.C., Miller, T.R., Strakhova, M.I., Vortherms, T.A., Wakefield, B.D., Wetter, J.M., Witte, D.G., Honore, P., Esbenshade, T.A., Brioni, J.D., Cowart, M.D., 2008. cis-4-(Piperazin-1-yl)-5,6,7a,8,9,10,11,11a-octahydrobenzofuro[2,3-h]quinazolin-2-amine (A-987306), a new histamine H4R antagonist that blocks pain responses against carrageenan-induced hyperalgesia. *Journal of Medicinal Chemistry* 51, 7094–7098.
- Loew, E.R., MacMillan, R., Kaiser, M.E., 1946. The anti-histamine properties of benadryl, beta-di-methylaminoethyl benzhydryl ether hydrochloride. *The Journal of Pharmacology and Experimental Therapeutics* 86, 229–238.
- Lucero, M.L., Gonzalo, A., Ganza, A., Leal, N., Soengas, I., Ioja, E., Gedey, S., Jahic, M., Bednarczyk, D., 2012. Interactions of bilastine, a new oral H(1) antihistamine, with human transporter systems. *Drug and Chemical Toxicology* 35 (Supplement 1), 8–17.
- Matsushita, A., Seike, M., Okawa, H., Kadawaki, Y., Ohtsu, H., 2012. Advantages of histamine H4 receptor antagonist usage with H1 receptor antagonist for the treatment of murine allergic contact dermatitis. *Experimental Dermatology* 21, 714–715.
- Maurer, M., Opitz, M., Henz, B.M., Paus, R., 1997. The mast cell products histamine and serotonin stimulate and TNF-alpha inhibits the proliferation of murine epidermal keratinocytes in situ. *Journal of Dermatological Science* 16, 79–84.
- Maurer, M., Weller, K., Bindslev-Jensen, C., Gimenez-Arnau, A., Bousquet, P.J., Bousquet, J., Canonica, G.W., Church, M.K., Godse, K.V., Grattan, C.E., Greaves, M.W., Hide, M., Kalogeromitros, D., Kaplan, A.P., Saini, S.S., Zhu, X.J., Zuberbier, T., 2011. Unmet clinical needs in chronic spontaneous urticaria. A GA(2)LEN task force report. *Allergy* 66, 317–330.
- McLeod, R.L., Mingo, G.G., Kreutner, W., Hey, J.A., 2005. Effect of combined histamine H1 and H3 receptor blockade on cutaneous microvascular permeability elicited by compound 48/80. *Life Sciences* 76, 1787–1794.
- Meiler, F., Zumkehr, J., Klunker, S., Ruckert, B., Akdis, C.A., Akdis, M., 2008. In vivo switch to IL-10-secreting T regulatory cells in high dose allergen exposure. *The Journal of Experimental Medicine* 205, 2887–2898.
- Metz, M., Maurer, M., 2011. Rupatadine for the treatment of allergic rhinitis and urticaria. *Expert Review of Clinical Immunology* 7, 15–20.
- Mirzahosseini, A., Dalmadi, B., Csutora, P., 2013. Histamine receptor H4 regulates mast cell degranulation and IgE induced FcεR1 upregulation in murine bone marrow-derived mast cells. *Cellular Immunology* 283, 38–44.
- Miura, M., Uno, T., 2010. Clinical pharmacokinetics of fexofenadine enantiomers. *Expert Opinion on Drug Metabolism & Toxicology* 6, 69–74.
- Miyano, K., Matsushita, S., Tsuchida, T., Nakamura, K., 2016. Inhibitory effect of a histamine 4 receptor antagonist on CCL17 and CCL22 production by monocyte-derived Langerhans cells in patients with atopic dermatitis. *The Journal of Dermatology* 43, 1024–1029.
- Molimard, M., Diquet, B., Benedetti, M.S., 2004. Comparison of pharmacokinetics and metabolism of desloratadine, fexofenadine, levocetirizine and mizolastine in humans. *Fundamental & Clinical Pharmacology* 18, 399–411.
- Mommert, S., Gschwandtner, M., Koether, B., Gutzmer, R., Werfel, T., 2012. Human memory Th17 cells express a functional histamine H4 receptor. *The American Journal of Pathology* 180, 177–185.
- Monroe, E.W., 1996. Safety and efficacy of loratadine in urticaria. *International Journal of Dermatology* 35, 837–841.
- Montoro, J., Mullol, J., Davila, I., Ferrer, M., Sastre, J., Bartra, J., Jauregui, I., Cuvillo, D.E.L., A. & Valero, A., 2011. Bilastine and the central nervous system. *Journal of Investigational Allergology & Clinical Immunology* 21 (Supplement 3), 9–15.
- Mullol, J., Bousquet, J., Bachert, C., Canonica, W.G., Gimenez-Arnau, A., Kowalski, M.L., Marti-Guadano, E., Maurer, M., Picado, C., Scadding, G., Van Cauwenberge, P., 2008. Rupatadine in allergic rhinitis and chronic urticaria. *Allergy* 63 (Supplement 87), 5–28.
- Murata, Y., Song, M., Kikuchi, H., Hisamichi, K., Xu, X.L., Greenspan, A., Kato, M., Chiou, C.-F., Kato, T., Guzzo, C., Thurmond, R.L., Ohtsuki, M., Furue, M., 2015. Phase 2a, randomized, double-blind, placebo-controlled, multicenter, parallel-group study of a H4 R-antagonist (JNJ-39758979) in Japanese adults with moderate atopic dermatitis. *The Journal of Dermatology* 42, 129–139.
- Nagarajan, G., Thangam, E.B., 2020. Effect of H4R antagonist N-(2-Aminoethyl)-5-Chloro-1H-indole-2-carboxamide (compound a) in a mouse model of allergic asthma. *Immunological Investigations* 1-14.
- Nagarajan, G., Mariappanadar, V., Tamizh, M., Kallappan, I., Elden, B.T., 2017. Effect of H4R antagonist N-(2-aminoethyl)-5-chloro-1H-indol-2-carboxamides and 5-chloro-2-(piperazin-1-ylmethyl)-1H-benzimidazole on histamine and 4-methylhistamine-induced mast cell response. *Journal of Receptor and Signal Transduction Research* 37, 304–313.
- Nakamura, T., Itadani, H., Hidaka, Y., Ohta, M., Tanaka, K., 2000. Molecular cloning and characterization of a new human histamine receptor, HH4R. *Biochemical and Biophysical Research Communications* 279, 615–620.
- Nettis, E., Calogiuri, G.F., Di Leo, E., Cardinale, F., Macchia, L., Ferrannini, A., Vacca, A., 2008. Once daily levocetirizine for the treatment of allergic rhinitis and chronic idiopathic urticaria. *Journal of Asthma and Allergy* 2, 17–23.
- Nettis, E., Delle Donne, P., Di Leo, E., Calogiuri, G.F., Ferrannini, A., Vacca, A., 2013. Rupatadine for the treatment of urticaria. *Expert Opinion on Pharmacotherapy* 14, 1807–1813.
- Neumann, D., Beermann, S., Burhenne, H., Glage, S., Hartwig, C., Seifert, R., 2013. The dual H3/4R antagonist thioperamide does not fully mimic the effects of the 'standard' H4R antagonist JNJ 7777120 in experimental murine asthma. *Naunyn-Schmiedeberg's Archives of Pharmacology* 386, 983–990.
- Nicoud, M.B., Formoso, K., Medina, V.A., 2019. Pathophysiological role of histamine H4 receptor in cancer: Therapeutic implications. *Frontiers in Pharmacology* 10, 556.
- Oda, T., Morikawa, N., Saito, Y., Masuho, Y., Matsumoto, S., 2000. Molecular cloning and characterization of a novel type of histamine receptor preferentially expressed in leukocytes. *The Journal of Biological Chemistry* 275, 36781–36786.
- Ohsawa, Y., Hirasawa, N., 2012. The antagonism of histamine H1 and H4 receptors ameliorates chronic allergic dermatitis via anti-pruritic and anti-inflammatory effects in NC/Nga mice. *Allergy* 67, 1014–1022.
- Oсна, N., Elliott, K., Khan, M.M., 2001. Regulation of interleukin-10 secretion by histamine in TH2 cells and splenocytes. *International Immunopharmacology* 1, 85–96.
- Park, S.J., Kim, K.S., Kim, E.J., 2008. Blockade of HERG K+ channel by an antihistamine drug brompheniramine requires the channel binding within the S6 residue Y652 and F656. *Journal of Applied Toxicology* 28, 104–111.
- Peng, H., Wang, J., Ye, X.Y., Cheng, J., Huang, C.Z., Li, L.Y., Li, T.Y., Li, C.W., 2019. Histamine H4 receptor regulates IL-6 and INF-γ secretion in native monocytes from healthy subjects and patients with allergic rhinitis. *Clinical and Translational Allergy* 9, 49.
- Petersen, L.J., Church, M.K., Skov, P.S., 1997. Histamine is released in the wheal but not the flare following challenge of human skin in vivo: A microdialysis study. *Clinical and Experimental Allergy* 27, 284–295.
- Petremann, M., Gueguen, C., Delgado Betancourt, V., Wersinger, E., Dyhrfeld-Johnsen, J., 2020. Effect of the novel histamine H4 receptor antagonist SENS-111 on spontaneous nystagmus in a rat model of acute unilateral vestibular loss. *British Journal of Pharmacology* 177, 623–633.
- Pini, A., Grange, C., Veglia, E., Argenziano, M., Cavalli, R., Guasti, D., Calosi, L., Ghè, C., Solarino, R., Thurmond, R.L., Camussi, G., Chazot, P.L., Rosa, A.C., 2018. Histamine H4 receptor antagonism prevents the progression of diabetic nephropathy in male DBA2/J mice. *Pharmacological Research* 128, 18–28.
- Polak, S., Wisniowska, B., Brandys, J., 2009. Collation, assessment and analysis of literature in vitro data on hERG receptor blocking potency for subsequent modeling of drugs' cardiotoxic properties. *Journal of Applied Toxicology* 29, 183–206.

- Popielek-Barczyk, K., Łażewska, D., Latacz, G., Olejarz, A., Makuch, W., Stark, H., Kieć-Kononowicz, K., Mika, J., 2018. Antinociceptive effects of novel histamine H3 and H4 receptor antagonists and their influence on morphine analgesia of neuropathic pain in the mouse. *British Journal of Pharmacology* 175, 2897–2910.
- Popov, T.A., Dumitrascu, D., Bachvarova, A., Bocsan, C., Dimitrov, V., Church, M.K., 2006. A comparison of levocetirizine and desloratadine in the histamine-induced wheal and flare response in human skin in vivo. *Inflammation Research* 55, 241–244.
- Potter, P.C., Kapp, A., Maurer, M., Guillet, G., Jian, A.M., Hauptmann, P., Finlay, A.Y., 2009. Comparison of the efficacy of levocetirizine 5 mg and desloratadine 5 mg in chronic idiopathic urticaria patients. *Allergy* 64, 596–604.
- Purohit, A., Duvernelle, C., Melac, M., Pauli, G., Frossard, N., 2001. Twenty-four hours of activity of cetirizine and fexofenadine in the skin. *Annals of Allergy, Asthma & Immunology* 86, 387–392.
- Purohit, A., Melac, M., Pauli, G., Frossard, N., 2003. Twenty-four-hour activity and consistency of activity of levocetirizine and desloratadine in the skin. *British Journal of Clinical Pharmacology* 56, 388–394.
- Purohit, A., Melac, M., Pauli, G., Frossard, N., 2004. Comparative activity of cetirizine and desloratadine on histamine-induced wheal-and-flare responses during 24 hours. *Annals of Allergy, Asthma & Immunology* 92, 635–640.
- Ring, J., Hein, R., Gauger, A., 2001. Desloratadine in the treatment of chronic idiopathic urticaria. *Allergy* 56 (Supplement 65), 28–32.
- Rojas-Zamorano, J.A., Esqueda-Leon, E., Jimenez-Anguiano, A., Cintra-Mcglone, L., Mendoza Melendez, M.A., Velazquez Moctezuma, J., 2009. The H1 histamine receptor blocker, chlorpheniramine, completely prevents the increase in REM sleep induced by immobilization stress in rats. *Pharmacology, Biochemistry, and Behavior* 91, 291–294.
- Rosssbach, K., Wendorf, S., Sander, K., Stark, H., Gutzmer, R., Werfel, T., Kietzmann, M., Bäumer, W., 2009. Histamine H4 receptor antagonism reduces hapten-induced scratching behaviour but not inflammation. *Experimental Dermatology* 18, 57–63.
- Rosssbach, K., Schaper, K., Kloth, C., Gutzmer, R., Werfel, T., Kietzmann, M., Bäumer, W., 2016. Histamine H4 receptor knockout mice display reduced inflammation in a chronic model of atopic dermatitis. *Allergy* 71, 189–197.
- Roy, M., Dumaine, R., Brown, A.M., 1996. Herg, a primary human ventricular target of the nonsedating antihistamine terfenadine. *Circulation* 94, 817–823.
- Russell, T., Stoltz, M., Weir, S., 1998. Pharmacokinetics, pharmacodynamics, and tolerance of single- and multiple-dose fexofenadine hydrochloride in healthy male volunteers. *Clinical Pharmacology and Therapeutics* 64, 612–621.
- Salcedo, C., Pontes, C., Merlos, M., 2013. Is the H4 receptor a new drug target for allergies and asthma? *Frontiers in Bioscience (Elite Edition)* 5, 178–187.
- Sanna, M.D., Borgonetti, V., Masini, E., Galeotti, N., 2020. Histamine H4 receptor stimulation in the locus coeruleus attenuates neuropathic pain by promoting the coeruleospinal noradrenergic inhibitory pathway. *European Journal of Pharmacology* 868, 172859.
- Sastre, J., 2008. Ebastine in allergic rhinitis and chronic idiopathic urticaria. *Allergy* 63 (Supplement 89), 1–20.
- Scadding, G., 2008. Optimal management of nasal congestion caused by allergic rhinitis in children: Safety and efficacy of medical treatments. *Paediatric Drugs* 10, 151–162.
- Scadding, G.K., Church, M.K. (Eds.), 2006. *Rhinitis*. Mosby Elsevier, London.
- Schaefer, U., Schmitz, V., Schneider, A., Neugebauer, E., 1999. Histamine induced homologous and heterologous regulation of histamine receptor subtype mRNA expression in cultured endothelial cells. *Shock* 12, 309–315.
- Schinkel, A.H., 1999. P-Glycoprotein, a gatekeeper in the blood-brain barrier. *Advanced Drug Delivery Reviews* 36, 179–194.
- Schirmer, B., Rother, T., Bruesch, I., Bleich, A., Werlein, C., Jonigk, D., Seifert, R., Neumann, D., 2020. Genetic deficiency of the histamine H4-receptor reduces experimental colorectal carcinogenesis in mice. *Cancers* 12, 4.
- Schwartz, J.C., Pollard, H., Quach, T.T., 1980. Histamine as a neurotransmitter in mammalian brain: Neurochemical evidence. *Journal of Neurochemistry* 35, 26–33.
- Seelig, A., Landwojtowicz, E., 2000. Structure-activity relationship of P-glycoprotein substrates and modifiers. *European Journal of Pharmaceutical Sciences* 12, 31–40.
- Seifert, R., Strasser, A., Schneider, E.H., Neumann, D., Dove, S., Buschauer, A., 2013. Molecular and cellular analysis of human histamine receptor subtypes. *Trends in Pharmacological Sciences* 34, 33–58.
- Seike, M., Furuya, K., Omura, M., Hamada-Watanabe, K., Matsushita, A., Ohtsu, H., 2010. Histamine H(4) receptor antagonist ameliorates chronic allergic contact dermatitis induced by repeated challenge. *Allergy* 65, 319–326.
- Shimizu, M., Fuse, K., Okudaira, K., Nishigaki, R., Maeda, K., Kusuhara, H., Sugiyama, Y., 2005. Contribution of OATP (organic anion-transporting polypeptide) family transporters to the hepatic uptake of fexofenadine in humans. *Drug Metabolism and Disposition* 33, 1477–1481.
- Shin, N., Covington, M., Bian, D., Zhuo, J., Bowman, K., Li, Y., Soloviev, M., Qian, D.-Q., Feldman, P., Leffet, L., He, X., He Wang, K., Krug, K., Bell, D., Czerniak, P., Hu, Z., Zhao, H., Zhang, J., Yeleswaram, S., Yao, W., Newton, R., Scherle, P., 2012. INCB38579, a novel and potent histamine H₄ receptor small molecule antagonist with anti-inflammatory pain and anti-pruritic functions. *European Journal of Pharmacology* 675, 47–56.
- Shiraiishi, Y., Jia, Y., Domenico, J., Joetham, A., Karasuyama, H., Takeda, K., Gelfand, E.W., 2013. Sequential engagement of FcεRI on mast cells and basophil histamine H(4) receptor and FcεRI in allergic rhinitis. *Journal of Immunology* 190, 539–548.
- Simons, F.E., 1994. H1-receptor antagonists. Comparative tolerability and safety. *Drug Safety* 10, 350–380.
- Simons, F.E., 1999a. H1-receptor antagonists: Safety issues. *Annals of Allergy, Asthma & Immunology* 83, 481–488.
- Simons, F.E., 1999b. Is antihistamine (H1-receptor antagonist) therapy useful in clinical asthma? *Clinical and Experimental Allergy* 29 (Supplement 3), 98–104.
- Simons, F.E., 1999c. Mizolastine: Antihistaminic activity from preclinical data to clinical evaluation. *Clinical and Experimental Allergy* 29 (Supplement 1), 3–8.
- Simons, F.E., 2004. Advances in H1-antihistamines. *The New England Journal of Medicine* 351, 2203–2217.
- Simons, F.E., Fraser, T.G., Reggin, J.D., Simons, K.J., 1995. Individual differences in central nervous system response to antihistamines (H1-receptor antagonists). *Annals of Allergy, Asthma & Immunology* 75, 507–514.
- Simons, F.E., Prenner, B.M., Finn Jr., A., Desloratadine Study Group, 2003. Efficacy and safety of desloratadine in the treatment of perennial allergic rhinitis. *The Journal of Allergy and Clinical Immunology* 111, 617–622.
- Simons, K.J., Benedetti, M.S., Simons, F.E., Gillard, M., Baltes, E., 2007. Relevance of H1-receptor occupancy to H1-antihistamine dosing in children. *The Journal of Allergy and Clinical Immunology* 119, 1551–1554.
- Simpson, K., Jarvis, B., 2000. Fexofenadine: A review of its use in the management of seasonal allergic rhinitis and chronic idiopathic urticaria. *Drugs* 59, 301–321.
- Singh, M., Jadhav, H.R., 2013. Histamine H3 receptor function and ligands: Recent developments. *Mini Reviews in Medicinal Chemistry* 13, 47–57.
- Singh-Franco, D., Ghin, H.L., Robles, G.I., Borja-Hart, N., Perez, A., 2009. Levocetirizine for the treatment of allergic rhinitis and chronic idiopathic urticaria in adults and children. *Clinical Therapeutics* 31, 1664–1687.
- Smit, M.J., Leurs, R., Alewijnse, A.E., Blauw, J., Van Nieu Amerongen, G.P., Van De Vrede, Y., Roovers, E., Timmerman, H., 1996. Inverse agonism of histamine H2 antagonist accounts for upregulation of spontaneously active histamine H2 receptors. *Proceedings of the National Academy of Sciences of the United States of America* 93, 6802–6807.
- Smits, R.A., Leurs, R., De Esch, I.J., 2009. Major advances in the development of histamine H4 receptor ligands. *Drug Discovery Today* 14, 745–753.
- Soldovieri, M.V., Miceli, F., Tagliatalata, M., 2008. Cardiotoxic effects of antihistamines: From basics to clinics (...and back). *Chemical Research in Toxicology* 21, 997–1004.
- Somma, T., Cinci, L., Formicola, G., Pini, A., Thurmond, R., Ennis, M., Bani, D., Masini, E., 2013. A selective antagonist of histamine H4 receptors prevents antigen-induced airway inflammation and bronchoconstriction in guinea pigs: Involvement of lipocortin-1. *British Journal of Pharmacology* 170, 200–213.
- Spencer, C.M., Faulds, D., Peters, D.H., 1993. Cetirizine. A reappraisal of its pharmacological properties and therapeutic use in selected allergic disorders. *Drugs* 46, 1055–1080.
- Staevska, M., Popov, T.A., Kralimarkova, T., Lazarova, C., Kraeva, S., Popova, D., Church, D.S., Dimitrov, V., Church, M.K., 2010. The effectiveness of levocetirizine and desloratadine in up to 4 times conventional doses in difficult-to-treat urticaria. *The Journal of Allergy and Clinical Immunology* 125, 676–682.
- Staub, A.M., 1937. Action de la thymoxyethyl-diethylamine (929F) et des éthers phénoliques sur le choc anaphylactique. *Comptes Rendus de la Société de Biologie* 125, 818–821.
- Stokes, J.R., Romero Jr., F.A., Allan, R.J., Phillips, P.G., Hackman, F., Misfeldt, J., Casale, T.B., 2012. The effects of an H3 receptor antagonist (PF-03654746) with fexofenadine on reducing allergic rhinitis symptoms. *Journal of Allergy Clinical Immunology* 129, 409–412, 412 e1–2.

- Strakhova, M.I., Cuff, C.A., Manelli, A.M., Carr, T.L., Witte, D.G., Baranowski, J.L., Vortherms, T.A., Miller, T.R., Rundell, L., Mcpherson, M.J., Adair, R.M., Brito, A.A., Bettencourt, B.M., Yao, B.B., Wetter, J.M., Marsh, K.C., Liu, H., Cowart, M.D., Brioni, J.D., Esbenshade, T.A., 2009. In vitro and in vivo characterization of A-940894: A potent histamine H4 receptor antagonist with anti-inflammatory properties. *British Journal of Pharmacology* 157, 44–54.
- Suessbrich, H., Waldegger, S., Lang, F., Busch, A.E., 1996. Blockade of HERG channels expressed in *Xenopus oocytes* by the histamine receptor antagonists terfenadine and astemizole. *FEBS Letters* 385, 77–80.
- Suwa, E., Yamaura, K., Oda, M., Namiki, T., Ueno, K., 2011. Histamine H(4) receptor antagonist reduces dermal inflammation and pruritus in a hapten-induced experimental model. *European Journal of Pharmacology* 667, 383–388.
- Suzuki, S., Takeuchi, K., Majima, Y., 2008. Localization and function of histamine H3 receptor in the nasal mucosa. *Clinical and Experimental Allergy* 38, 1476–1482.
- Tagliatalata, M., Castaldo, P., Pannaccione, A., Giorgio, G., Annunziato, L., 1998. Human ether-a-gogo related gene (HERG) K⁺ channels as pharmacological targets: Present and future implications. *Biochemical Pharmacology* 55, 1741–1746.
- Tagliatalata, M., Castaldo, P., Pannaccione, A., Giorgio, G., Genovese, A., Marone, G., Annunziato, L., 1999. Cardiac ion channels and antihistamines: Possible mechanisms of cardiotoxicity. *Clinical and Experimental Allergy* 29 (Supplement 3), 182–189.
- Tagliatalata, M., Timmerman, H., Annunziato, L., 2000. Cardiotoxic potential and CNS effects of first-generation antihistamines. *Trends in Pharmacological Sciences* 21, 52–56.
- Takahashi, Y., Kagawa, Y., Izawa, K., Ono, R., Akagi, M., Kamei, C., 2009. Effect of histamine H4 receptor antagonist on allergic rhinitis in mice. *International Immunopharmacology* 9, 734–738.
- Tannenbaum, C., Paquette, A., Hilmer, S., Holroyd-Leduc, J., Carnahan, R., 2012. A systematic review of amnesic and non-amnesic mild cognitive impairment induced by anticholinergic, antihistamine, GABAergic and opioid drugs. *Drugs & Aging* 29, 639–658.
- Tannergren, C., Petri, N., Knutson, L., Hedeland, M., Bondesson, U., Lennernas, H., 2003. Multiple transport mechanisms involved in the intestinal absorption and first-pass extraction of fexofenadine. *Clinical Pharmacology and Therapeutics* 74, 423–436.
- Ten Eick, A.P., Blumer, J.L., Reed, M.D., 2001. Safety of antihistamines in children. *Drug Safety* 24, 119–147.
- Teuscher, C., Subramanian, M., Noubade, R., Gao, J.F., Offner, H., Zachary, J.F., Blankenhorn, E.P., 2007. Central histamine H3 receptor signaling negatively regulates susceptibility to autoimmune inflammatory disease of the CNS. *Proceedings of the National Academy of Sciences of the United States of America* 104, 10146–10151.
- Thangam, E.B., Jemima, E.A., Singh, H., Baig, M.S., Khan, M., Mathias, C.B., Church, M.K., Saluja, R., 2018. The role of histamine and histamine receptors in mast cell-mediated allergy and inflammation: The hunt for new therapeutic targets. *Frontiers in Immunology* 9, 1873.
- Theoharides, T.C., 1989. Histamine2 (H2)-receptor antagonists in the treatment of urticaria. *Drugs* 37, 345–355.
- Thurmond, R.L., 2015. The histamine H4 receptor: From orphan to the clinic. *Frontiers in Pharmacology* 6, 65.
- Thurmond, R.L., Desai, P.J., Dunford, P.J., Fung-Leung, W.-P., Hofstra, C.L., Jiang, W., Nguyen, S., Riley, J.P., Sun, S., Williams, K.N., Edwards, J.P., Karlsson, L., 2004. A potent and selective histamine H4 receptor antagonist with anti-inflammatory properties. *The Journal of Pharmacology and Experimental Therapeutics* 309, 404–413.
- Thurmond, R.L., Chen, B., Dunford, P.J., Greenspan, A.J., Karlsson, L., La, D., Ward, P., Xu, X.L., 2014. Clinical and preclinical characterization of the histamine H(4) receptor antagonist JNJ-39758979. *The Journal of Pharmacology and Experimental Therapeutics* 349, 176–184.
- Thurmond, R.L., Venable, J., Savall, B., La, D., Snook, S., Dunford, P.J., Edwards, J.P., 2017. Clinical development of histamine H4 receptor antagonists. In: *Handbook of Experimental Pharmacology*. Springer.
- Tiligada, E., 2012. Editorial: Is histamine the missing link in chronic inflammation? *Journal of Leukocyte Biology* 92, 4–6.
- Van Cauwenberge, P., De Belder, T., Sys, L., 2004. A review of the second-generation antihistamine ebastine for the treatment of allergic disorders. *Expert Opinion on Pharmacotherapy* 5, 1807–1813.
- Venail, F., Attali, P., Wersinger, E., Gomeni, R., Poli, S., Scherber, S., 2018. Safety, tolerability, pharmacokinetics and pharmacokinetic-pharmacodynamic modelling of the novel H4 receptor inhibitor SENS-111 using a modified calorimetric test in healthy subjects. *British Journal of Clinical Pharmacology* 84, 2836–2848.
- Vuurman, E.F., Van Veggel, L.M., Uiterwijk, M.M., Leutner, D., O'Hanlon, J.F., 1993. Seasonal allergic rhinitis and antihistamine effects on children's learning. *Annals of Allergy* 71, 121–126.
- Vuurman, E.F., Van Veggel, L.M., Sanders, R.L., Muntjewerff, N.D., O'Hanlon, J.F., 1996. Effects of semprex-D and diphenhydramine on learning in young adults with seasonal allergic rhinitis. *Annals of Allergy, Asthma & Immunology* 76, 247–252.
- Walker, S., Khan-Wasti, S., Fletcher, M., Cullinan, P., Harris, J., Sheikh, A., 2007. Seasonal allergic rhinitis is associated with a detrimental effect on examination performance in United Kingdom teenagers: Case-control study. *The Journal of Allergy and Clinical Immunology* 120, 381–387.
- Wang, M., Han, J., Domenico, J., Shin, Y.S., Jia, Y., Gelfand, E.W., 2016. Combined blockade of the histamine H1 and H4 receptor suppresses peanut-induced intestinal anaphylaxis by regulating dendritic cell function. *Allergy* 71, 1561–1574.
- Wang, W.W., Pan, Y.L., Yu, H.W., Zhang, B., Shao, S.W., 2019. Histamine H4 receptor regulates Th2-cytokine profile through thymic stromal lymphopoietin in allergic rhinitis. *European Archives of Oto-Rhino-Laryngology* 276, 1655–1661.
- Warren, R., Simpson, H., Hilchie, J., Cimbura, G., Lucas, D., Bennett, R., 1981. Drugs detected in fatally injured drivers in the province of Ontario. *Proc. Int. Counc. Alcohol Drugs Traffic Safety Conf.* 1981. International Council on Alcohol, drugs and Traffic Safety, pp. 203–217.
- Werfel, T., Layton, G., Yeadon, M., Whitlock, L., Osterloh, I., Jimenez, P., Liu, W., Lynch, V., Asher, A., Tsianakas, A., Purkins, L., 2019. Efficacy and safety of the histamine H4 receptor antagonist ZPL-3893787 in patients with atopic dermatitis. *The Journal of Allergy and Clinical Immunology* 143, 1830–18374.
- Wieland, K., Laak, A.M., Smit, M.J., Kuhne, R., Timmerman, H., Leurs, R., 1999. Mutational analysis of the antagonist-binding site of the histamine H(1) receptor. *The Journal of Biological Chemistry* 274, 29994–30000.
- Yamamoto, K., Okui, R., Yamatodani, A., 2019. Activation of orexinergic and histaminergic pathway involved in therapeutic effect of histamine H4 receptor antagonist against cisplatin-induced anorexia in mice. *Naunyn-Schmiedeberg's Archives of Pharmacology* 392, 925–936.
- Yamaura, K., Oda, M., Suwa, E., Suzuki, M., Sato, H., Ueno, K., 2009. Expression of histamine H4 receptor in human epidermal tissues and attenuation of experimental pruritus using H4 receptor antagonist. *The Journal of Toxicological Sciences* 34, 427–431.
- Yan, Z.-Q., Zhang, R.-X., Yu, S.-Q., Wen, W., Hong, J.-K., Zhang, J., Shao, X.-L., Gao, S.-H., 2010. Effect of histamine H4 receptor and its antagonist on allergic rhinitis in rats. *Zhonghua Er Bi Yan Hou Tou Jing Wai Ke Za Zhi* 45, 477–480.
- Yanai, K., 2012. Anticholinergic activity of antihistamines. *Clinical Neurophysiology* 123, 633–634.
- Yanai, K., Zhang, D., Tashiro, M., Yoshikawa, T., Naganuma, F., Harada, R., Nakamura, T., Shibuya, K., Okamura, N., 2011. Positron emission tomography evaluation of sedative properties of antihistamines. *Expert Opinion on Drug Safety* 10, 613–622.
- Yuan, S.F., Guan, J.Z., Hao, Y., Chai, J.Y., Gao, F.L., Shi, S.Q., Wang, J.X., He, J., Shi, J.H., 2017. Effectiveness and safety of the combination of the traditional Chinese medicine prescription jade screen and desloratadine in the treatment of chronic urticaria: A systematic review and meta-analysis of randomized controlled trials. *Evidence-based Complementary and Alternative Medicine* 2017, 1390301.
- Zhang, W., Zhang, X., Zhang, Y., Qu, C., Zhou, X., Zhang, S., 2019. Histamine induces microglia activation and the release of proinflammatory mediators in rat brain via H1R or H4R. *Journal of Neuroimmune Pharmacology* 15, 280–291.
- Zhou, P., Homborg, J.R., Fang, Q., Wang, J., Li, W., Meng, X., Shen, J., Luan, Y., Liao, P., Swaab, D.F., Shan, L., Liu, C., 2019. Histamine-4 receptor antagonist JNJ777120 inhibits pro-inflammatory microglia and prevents the progression of Parkinson-like pathology and behaviour in a rat model. *Brain, Behavior, and Immunity* 76, 61–73.
- Zuberbier, T., Aberer, W., Asero, R., Abdul Latiff, A.H., Baker, D., Ballmer-Weber, B., Bernstein, J.A., Bindslev-Jensen, C., Brzoza, Z., Buense Bedrikow, R., Canonica, G.W., Church, M.K., Craig, T., Danilycheva, I.V., Dressler, C., Ensina, L.F., Gimenez-Arnau, A., Godse, K., Goncalo, M., Grattan, C., Hebert, J., Hide, M., Kaplan, A., Kapp, A., Katelaris, C.H., Kocaturk, E., Kulthanan, K., Larenas-Linnemann, D., Leslie, T.A., Magerl, M., Mathelier-Fusade, P., Meshkova, R.Y., Metz, M., Nast, A., Nettis, E., Oude-Elberink, H., Rosumeck, S., Saini, S.S., Sanchez-Borges, M., Schmid-Grendelmeier, P., Staubach, P., Sussman, G., Toubi, E., Vena, G.A., Vestergaard, C., Wedi, B., Werner, R.N., Zhao, Z., Maurer, M., 2018. The EAACI/GA(2)LEN/EDF/WAO guideline for the definition, classification, diagnosis and management of urticaria. *Allergy* 73, 1393–1414.